Minireview

Dysregulation of multiple signaling pathways: A possible cause of cerebral palsy

Jyoti Upadhyay1, Mohd Nazam Ansari2 , Abdul Samad3 and Ashutosh Sayana4

1School of Health Sciences and Technology, University of Petroleum and Energy Studies, Dehradun 248007, India; 2Department of Pharmacology & Toxicology, College of Pharmacy, Prince Sattam Bin Abdulaziz University, Al-Kharj 11942, Saudi Arabia; ³Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Tishk International University, Erbil 44001, Kurdistan Region, Iraq; 4Government Doon Medical College, Dehradun 248001, India

Corresponding author: Mohd Nazam Ansari. Email: nazam.ansari@gmail.com

Impact Statement

Cerebral palsy (CP) is a permanent disability that is defined by brain function impairment that results in improper posture and aberrant movement patterns. Prematurity and hypoxic–ischemic damage are two well-known factors contributing to the development of CP. It has been discovered that the neuroinflammation caused by maternal cytokine response plays an important part in the pathogenesis of fetal inflammation response, which is one of the contributing causes for CP, and it persists even after the delivery of children with CP. The canonical Wnt signaling pathway is crucial for mammalian fetal brain development and it governs many processes such as neurogenesis. The antagonistic activity of GSK-3 in the Wnt signaling pathway plays an important role in neurogenesis, neural development, and brain development.

Abstract

Cerebral palsy (CP) is a lifelong disability characterized by the impairment of brain functions that result in improper posture and abnormal motor patterns. Understanding this brain abnormality and the role of genetic, epigenetic, and non-genetic factors such as signaling pathway dysregulation and cytokine dysregulation in the pathogenesis of CP is a complex process. Hypoxic–ischemic injury and prematurity are two well-known contributors of CP. Like in the case of other neurodevelopmental disorders such as intellectual disability and autism, the genomic constituents in CP are highly complex. The neuroinflammation that is triggered by maternal cytokine response plays a critical role in the pathogenesis of fetal inflammation response, which is one of the contributing factors of CP, and it continues even after the birth of children suffering from CP. Canonical Wnt signaling pathway is important for the development of mammalian fetal brain and it regulates distinct processes including neurogenesis. The glycogen synthase kinase-3 (GSK-3) antagonistic activity in the Wnt signaling pathway plays a crucial role in neurogenesis and neural development. In this review, we investigated several genetic and non-genetic pathways that are involved in the pathogenesis of CP and their regulation, impairment, and implications for causing CP during embryonic growth and developmental period. Investigating the role of these pathways help to develop novel therapeutic interventions and biomarkers for early diagnosis and

treatment. This review also helps us to comprehend the mechanical approach of various signaling pathways, as well as their consequences and relevance in the understanding of CP.

Keywords: Cerebral palsy, genetics, epigenetics, cytokine regulation, Wnt signaling pathway

Experimental Biology and Medicine **2022; 247: 779–787. DOI: 10.1177/15353702221081022**

Introduction

Cerebral palsy (CP) is a non-progressive neurodevelopmental disorder that affects approximately one in five hundred children. Its clinical features include impairment in movement and posture, motor deficiency, lack of coordination, and upper motor neuron disorder involving spasticity.¹ The motor impairment of CP is often accompanied by other related impairments such as epilepsy, intellectual disability, and sensory disorders. Although numerous risk factors associated with CP are identified, for many individuals, it is difficult to determine the etiology of CP since it involves multiple

hypoxic-ischemia, which accounts for less than 10% of cases, is the main cause of CP.^{2,3} The risk factors associated with CP include congenital malformations, multiple gestations, preterm birth, intrauterine inflammation and infection, birth asphyxia, thrombophilia, and perinatal stroke. An important pathophysiological mechanism that is observed is the infection of amniotic fluid and intra-amniotic inflammation that causes damage to the developing brain of the fetus, leading to CP. This damage may persist for many years. Several neurological and neuropsychiatric disorders are associated with perinatal infection and inflammation, and these are found

mechanisms. Some studies report that acute intrapartum

to have long-term consequences on the child's brain.⁴ Brain encephalopathy in term-born infants accounts for 24% of all cases and is reported to cause inflammatory responses in the first week after birth, which is correlated with the level of brain injury.5,6 Preterm deliveries affect approximately 15million births7 and are found to be important risk factors for many congenital disorders including cognitive impairment, CP, autism, and mental disorders that are discovered later in life.⁸

Several studies investigated that magnetic resonance imaging (MRI) of cerebral phenotypes of preterm babies are associated with later functions and involves dysregulation of neural pathways, structural alterations of cerebral cortex and gray matter, and diffused white matter diseases.⁹⁻¹² Many studies indicated that there has been a decline in the incidence rate of CP over the last few decades due to improved prenatal care and obstetric practice. These results led some researchers to investigate the "unknown pathophysiologic approaches" that account for significant proportion of CP.13 Out of the studies conducted, some of them suggested that the unknown pathophysiology of this disease may include genetic factors and dysregulation of some neuronal pathways. In this article, we investigated the association of genetic or epigenetic factors, neuronal signaling pathways, and inflammatory response alterations with CP. Understanding these pathways can aid in the development of novel, effective, and safe therapeutic treatments for CP.

Methodology

This study aimed to investigate the clear insights about the mechanistic pathways involved in the causation of CP as well as the food pattern and nutritional status of children affected with CP. First, we have identified the research articles with appropriate words and phrases. We have used search engines like PubMed, Google Scholar, and MEDLINE. The key phrases used were "cerebral palsy" and "signaling pathways involved in cerebral palsy" combined with "nutritional status" and "food intake in CP children." Total number of papers identified (*n*=500). The inclusion criteria for paper selection were that we have identified papers including studies related to multiple signaling pathways and nutritional status of children with CP. Total number of papers after inclusion–exclusion consideration (*n*=150) and full-text screened researched articles (*n*=100). Total number of papers included into the study (*n*=75).

Genetic or epigenetic implications in CP

Current research investigates that 30% of CP cases are caused by genetic or epigenetic factors.14–16 Four main types of DNA variations are identified that contribute to the pathogenesis of CP. The unfortunate outcome of the many mutations is that it results in the loss of important cellular functions of several proteins that are encoded by these genes. Figure 1 depicts the contribution of genetics in the pathophysiology of CP.

The effects of genetic mutation may vary depending on its nature, involvement of environmental factors, and the individual genomic profiles where mutation occurs. A mutation can either be severe, where the deleterious mutation

causes major effects, or minor, where the damage caused is less enough to not affect the protein function. A deleterious mutation is sufficient to cause CP in most individuals, while in others cases, minor mutations coupled with environmental factors like hypoxia may cross thresholds and cause severe neuromotor disorders. For instance, CP is not caused by one single mutation, but the combined effect of several less damaging gene mutations in a polygenic manner.17 A study conducted in hypoxic–ischemic rabbit model that is induced by aortic ligation revealed that when their litters were exposed to the same insult, they developed motor impairment suggesting that genetic or epigenetic modulators may mediate adverse motor outcome and extent of injury.18 Mitochondrial DNA is suggested to be a potential biomarker for many dysfunctions of mitochondria, and it is found to be related to many diseases including neurological diseases, traumatic brain injury, and autism. Lu *et al.*19 investigated the association of mitochondrial DNA copy number with CP and reported that a decline in mitochondrial DNA copy number implied mitochondrial dysfunction and CP. A study that was conducted reported that the mutation in mitochondrial DNA 8993 is associated with Leigh encephalopathy in children, where CP or other neurological impairment was found in other family members. These family members were examined to analyze the correlation between mutant mitochondrial DNA and the severity of the disease.²⁰ It was observed in 10–20% of the cases that genomic copy number variations, which involve deletions or duplications, may cause CP.21–23 Microarray techniques were used by researchers on a cohort of 52 subjects suffering from cryptogenic CP. In this study, the ACMG (American College of Medical Genetics and Genomics) standard was applied to interpret the results where it was identified that 31% of cases were pathogenic or likely to be pathogenic copy number variation.²² Another study used the same technique on 147 non-selected CP patients and found copy number changes that met ACMG requirements.23 Epigenomic studies revealed some alterations in axon structure, actin cytoskeleton, and cell signaling, which was aligned with genomic studies. Alterations caused by DNA methylation that show changes in mitogenactivated protein kinase (MAPK) signaling, inflammation, cytokine–cytokine receptors, and Ras signaling at birth were observed in monozygotic twins discordant for CP.24 In nonselected subjects with CP, there were differences identified in the methylation process in the actin cytoskeleton, axon structure, crosstalk between dendritic cells and natural killer cells, insulin receptors, neuregulin, transforming growth factor beta (TGF-β), Wnt signaling, and phosphoinositide 3-kinases (PI3Ks)/Akt signaling pathways.25 Many cellular signaling pathways that involve synaptic dysfunction, activity-dependent translation and transcription, neuroglia signaling, and neuroinflammation are dysregulated in numerous neurodevelopmental disorders, which necessitate in-depth research by utilizing animal models. It was seen that transcriptomic studies of the lymphoblastoid cell lines of CP subjects (*n*=182) who are exposed to genetic, environmental, and indeterminate causes reported 387 differentially expressed genes.26 An analysis of these gene pathways demonstrates a down-regulation of cell signaling and transduction including an upregulation of immune system-related

Figure 1. Genetic implications in cerebral palsy showing main sites and types. (A color version of this figure is available in the online journal.)

genes, brain-derived neurotrophic factor (bdnf), and altered amyloid precursor protein A. Some studies suggested an overlap in dysregulation of MAPK signaling and result of the epigenetic analysis. A report showed that defects in genes that regulate cell-signaling pathway – that is, MAPK, PI3K, or Akt – may cause CP and neuronal signaling defects.26

Implications of cytokine dysregulations in CP

While one of the etiologies behind the CP is neuroinflammation that is triggered by several mechanisms including maternal/fetal infection, hypoxia, maternal preeclampsia, and stroke, the other possibility is genetic etiologies.27 The cytokine response that is observed in a maternal amniotic fluid includes increased levels of interleukins (IL-6 and IL-1β) and tumor necrosis factor (TNF) alpha. In the fetal compartment, the level of IL-6 is found to play an important role in the pathogenesis of inflammatory syndrome in developing fetuses, which is associated with CP. This inflammatory pathway appears to continue even after the birth of CP subjects and gives rise to the "sustained inflammation hypothesis" that suggests that prenatal, antenatal, or neonatal pro-inflammatory cytokines induce inflammation that contributes to dysregulation of cytokine pathways.28 A study conducted on school children having post-neonatal encephalopathy (NE) explored the cytokine response to report an abnormally elevated level of cytokines in these children. The level of granulocyte-monocyte-cerebrospinal fluid (GM-CSF), IL-6, IL-8, and TNF- β was found to be significantly high in children with NE when compared to control

subjects. IL-8 and GM-CSF were found to be significantly elevated in children with NE upon stimulation with LPS (lipopolysaccharides) when compared with age-matched controls. Hypo-responsiveness of LPS in various cytokines among schoolchildren demonstrates an altered immune response.29 A study conducted by Huang *et al.*30 investigated the association of umbilical cord blood cytokines with CP in premature babies. They performed enzyme-linked immunoassay technique and identified a significantly high level of IL-8, PGE2, and myeloperoxidase (MPO) level in preterm babies with a gestation period of 32weeks when compared to full-term babies. These cytokines are not related to gestational age but to preterm birth. It was also seen that cytokine IL-8 was increased in CP-affected preterm infants but not MPO. Some of the evidence suggested that preterm deliveries caused by cytokine induction are mostly due to chorioamnionitis. It was observed that intrauterine inflammation or infection that caused activation of cytokines and elevated the level of pro-inflammatory cytokines in amniotic fluid and neonatal blood that the preterm baby was exposed to was identified as an important reason for preterm deliveries, CP, and periventricular leukomalacia (Figure 2).³¹ One study evaluated the association of four TNF-α promoter single nucleotide polymorphism (SNPs), two IL1β SNPs, and one IL-6 polymorphism that is susceptible to CP in preterm babies. Their results investigated IL-1β and TNF- $α$ polymorphism that is related to a higher level of cytokines in the circulation, to find their role in genetic susceptibility to damage white matter and cause CP in preterm infants.³² Animal model studies provided evidence that ischemic injury and inflammation/infection-induced brain injury play a major

Figure 2. Association of cytokines, reactive oxygen species, and maternal infection and inflammation that causes microglial activation to the prematurity and periventricular leukomalacia that are responsible for cerebral palsy. (A color version of this figure is available in the online journal.)

role in CP pathogenesis. A study that was based on reverse transcriptase polymerase chain reaction (PCR) methods showed elevated levels of pro-inflammatory cytokines including IL-6, IL-1β, and MCP-1 (monocyte chemoattractant protein-1) in the brains of mouse pups that were exposed to in-utero lipopolysaccharides (LPS).³³ Some studies revealed that a dose-dependent elevation was observed in the expression of TNF- $α$ and IL-1β mRNA in rat fetal brain that are exposed to in-utero LPS. In addition to this, the hippocampus and cortical region of the brain observed a significant decrease in the level of myelin basic protein, elevations in the level of glial protein (acidic or fibrillar), positive astrocytes, and changes in immune reactivity of oligodendrocytes (OLs).34 Table 1 represents the association of inflammatory cytokines with CP.35–48

Some studies reported that immune abnormalities have a strong relationship with CP and erythropoietin (EPO) plays a neuroprotective role in cell injuries associated with CP. EPO is a glycoprotein containing 165 amino acids, and is known as pleiotropic cytokine. Some studies suggested that EPO performs some non-hematopoietic actions like protection, maintenance, and development of nervous system. EPO in brain is normally secreted from astrocytes and EPO receptors are expressed principally in neurons. EPO acts as neuroprotective as well as anti-inflammatory agent by activating janus kinase 2 (JAK2)/signal transducers and activators of transcription 5 (STAT5) pathway. EPO binds with erythropoietin receptor (EPOR) in the extracellular domain causes conformational changes of receptor homodimers and results

in rapid phosphorylation of tyrosine residues of JAK2 and its activation in turn recognized by several Src homology-2 (SH-2) domain containing signal transduction molecules involving STAT5. Activation of this pathway in turn activates p85 subunit of PI3PK, NF-β, MAPK, and STAT5. STAT5 phosphorylation causes translocation and dimerization of the nucleus by acting as transcription factor and regulates the expression of various EPO-responsive genes. STAT5 is dephosphorylated by intra nuclear tyrosine phosphatase, and thus terminates the process of signal transduction. Thus, EPO provides neuroprotection by activating JAK/STAT pathway. Hypoxia inducible factor-1 (HIF-1) activated during inflammation enhances the secretion of EPO. It plays the role of anti-inflammatory agent by (1) inhibiting the expression of IL-1 and IL-6 induced by ischemia; (2) stimulating inflammatory cell death by pathways including phosphatidylserine exposure, protein kinase B, and activated microglia; (3) reducing oxidative stress, inflammatory response and myelin basic protein during immune reaction.⁴⁹ The cytokine profile study in postnatal childhood revealed key mediators of cell injury in CP and provided a better understanding of its pathophysiology that help to develop novel therapeutic interventions.

Implications of PI3K–Akt–Wnt pathway in CP

The Wnt signaling pathway is important for cell patterning, regulation of stem cells, and cell cycle during the mammalian

IL: interleukin; CP: cerebral palsy; LPS: lipopolysaccharides; TNF: tumor necrosis factor; CNS: central nervous system; VEGF: vascular endothelial growth factor.

fetal growth and development including the brain.50,51 Among adults, the Wnt pathways affect the regeneration and regulation of many tissues by homeostasis and proliferation of stem cells.52 It also maintains critical processes like axon remodeling, neuritic outgrowth, neurogenesis, and synaptic plasticity. Canonical Wnt signaling pathway suppresses glycogen synthase kinase-3 (GSK-3) and stimulates downstream regulation of signaling. While inhibiting the functions of canonical pathway of Wnt,53,54 GSK-3 also regulate other pathways that are involved in the development and function of neurons. Figure 3 represents the GSK-3 regulation of the Wnt and Akt signaling pathways, neurotrophic growth factor that activates Akt signaling to phosphorylate GSK-3 and suppresses it by allowing downstream effectors activation to promote cell survival. Neurogenesis is promoted by Wnt genes that are activated by β-catenin stabilization in Axin complex that is caused by the inhibition of GSK-3 by Wnts. It is observed that lithium antagonizes GSK-3 pools in both Wnt and Akt signaling and activate the pathway to function. Therefore, lithium activates the process of neurogenesis through Wnt/β-catenin signaling activation, and thus enhances cell survival. In the absence of Wnt ligands, GSK-3, APC (adenomatous polyposis coli), which is a key tumor suppressor gene, and β-catenin, which is a transcriptional co-activator, bind directly to the Axin protein complex to facilitate the β-catenin phosphorylation by GSK-3 that causes proteasome-dependent degradation by targeting β-catenin. Binding Wnt ligands to frizzled receptors activates the phosphorylation of LRP5 (low-density lipoprotein receptor-related protein-5 and 6) and co-receptors that cause inhibition of GSK-3 and stabilization of β-catenin. The stabilized form of β-catenin enters the nucleus and interacts with transcription factors like LEF (lymphocyte enhancer factor or T-cell factor [TCF]) family to stimulate transcription.55

Not only does GSK-3 regulate the Wnt signaling pathway, but it also regulates other signaling pathways including growth factor, notch, and sonic hedgehog pathway through Akt, thereby affecting the cell survival in the brain.51 Neutrophils and growth factors, along with insulin, stimulate Akt and PI3K (phosphatidylinositol-3-kinase). This causes phosphorylation of GSK-3 at *N*-terminal of serine residues (GSK-3β at Ser9 and GSK-3α at Ser21) to form pseudosubstrate motif that inhibits GSK-3 and allows downstream effector activation such as mTOR (mammalian target of rapamycin) and glycogen synthase.⁵⁶ Significantly, it was suggested that GSK-3 in Axin complex of Wnt pathway was not regulated by the phosphorylation of serine residues at *N*-terminal, Wnt ligands, which were neither neurotrophin nor insulin/Akt, that induced phosphorylation of Ser9 or 21 of GSK-3 was associated with Axin complex.⁵⁷ In addition to this, the GSK-3 pool associated with Akt and Wnt signaling responses was regulated by distinct mechanisms and subcellular pools that were demonstrated in double knock-in mice, where the phosphorylation of GSK-3bser9 and GSK-3aser21 sites was mutated to alanine.58

During embryonic development, Wnt signaling plays a critical role in the development of neural tube. It suppresses the anterior and promotes the posterior development of the neural tubes. Thus, while the inhibition of Wnt signaling pathway causes a reduced posterior development and enhanced anterior development, aberrant Wnt pathway stimulation causes reduced anterior development and enhanced posterior.59 In continuation with this process, Wnt antagonists such as DKK1 cause anterior localization and is important for the anterior development of neural tube.^{59,60} In the later stages of development, Wnt signaling causes patterning of neural tube by generating signal centers in the hindbrain including creating a midbrain–hindbrain

Figure 3. GSK-3 regulations of Wnt and Akt signaling pathways. (A color version of this figure is available in the online journal.)

boundary and restricting rhombomere boundaries.59–61 Wnt signaling is also important for neural tube patterning in dorsal or ventral form. Wnt1 and Wnt3a expressions occur in the dorsal neural tube, and if they are deleted, the ventral cell will expand at the cost of dorsal fates.⁶² Overexpression of genes *Wnt1* or *Wnt3a* causes extension of dorsal fates.62 Wnt signaling is important for neural crest specification since it promotes cell fate in dorsal and suppresses ventral cell fates in telencephalon during embryonic development.⁵⁹

The studies showed that Wnt signaling was important for the proliferation of precursor cells of a neuron during brain development. It was observed in a study that during the development of chick neural tube, overexpression of Wnt1 and Wnt3a, along with β-catenin stabilization, caused an increase in proliferation of neural precursor cells, whereas negative expression of dominant TCF4 resulted in decreased cell proliferation.63 Another study in mice indicated that overexpression of Wnt1 induced neuronal cell proliferation and size expansion in the caudal midbrain area that resulted in substantial midbrain enlargement.⁶⁴ In addition, the loss of β-catenin function in the mesencephalon, diencephalon, and hindbrain caused a decrease in the size of the midbrain by reducing the progenitor cell domain, while gain of β-catenin function caused an increase in brain size by expanding the progenitor cell domain.65 The loss of Wnt3 function caused a reduced proliferation of hippocampal neural progenitor cells and interrupted the development of the hippocampus region.66 A similar type of defect was observed in the dorsal telencephalon when β-catenin was deleted.⁶⁷ All these studies suggested that the Wnt/β-catenin signaling process stimulated progenitor cellular proliferation for the development of neural tube, midbrain, and hippocampus during the fetal growth development period.

The research deduced that apart from patterning and cellular proliferation, Wnt signaling promoted neurite development and caused an impact on axonal size, branching, remodeling, and complexity.68 It was observed that Wnt7a expression in cultured granular cells of the cerebellum caused an increase in axon size, neurite growth, and growth cone size while earlier studies observed that Wnt antagonist secreting frizzled receptor associated protein caused axonal remodeling and reduced growth cone size.^{69,70} Some of the studies also suggested that GSK-3 inhibitors were involved in promoting neurite size, outgrowth, and formation of an axon, axon size, and branching in several different cell types including granular cells of the cerebellum, neurons of dorsal root ganglion, and hippocampus.69–71 It was also observed that the Wnt in spinal cord development maintained the direction of commissural axon after it passed through the midline. This activity relied upon the aPKC (atypical protein kinase) and PI3K, but not on LTP6, which indicated non-involvement of canonical Wnt/β-catenin pathway.72 Moreover, it was deduced from the study that Wnt and β-catenin facilitated the growth of dendrites and their branching, synaptic formation, and plasticity in the cerebellum.53,73 These studies suggested the critical role of the Wnt signaling pathway in mammalian brain development including neurogenesis. Hence, it was deduced that any dysregulation of these pathways could cause several neuro-impairment disorders like CP, autism, and others. Therefore, for developing any novel therapeutic interventions for the treatment of these disorders, further research was required in this area.

Nutritional status and food pattern of children with CP

Association of CP with comorbidities like malnutrition, gastrointestinal (GI) symptoms, impaired growth and development, epilepsy, intellectual disability was observed in

many case studies.74,75 Motor dysfunction in CP causes oropharyngeal dysphagia. This may reduce food intake and consequently causes malnutrition, lung infection and pulmonary aspiration.76 One study reported that many children with CP considered "taste" to be very important in their meals, other hemiplegic group suggested "nutrition" to be the most important. Paraplegic children and children with severe brain injury usually preferred "sweet taste" while quadriplegic children preferred greasy taste. Children with CP engaged in very few outdoor activities; therefore, they have poor synthesis of vitamin D, and intake of calcium and vitamin D is important.77 Decreased food intake along with anticonvulsant medications causes decline in the level of bone mineral density. This results in functional impairment, muscular weaknesses, and pathological bone fractures. Inadequate food intake causes macro- and micro-nutrient deficiencies, especially anemia, vitamin A, and vitamin B complex deficiencies. The "Quality of Life" of the children with CP and their caregivers were suboptimum; correction of nutritional deficiencies especially vitamins A, D and B complex, anemia, and mental and physical support is suggested for the well-being of children suffering from CP.78

Future perspectives and conclusion

Genetic etiologies are found to be an important contributor to the development of CP, particularly through impaired brain development and dysregulation of several signaling pathways in response to associated risk factors. The number of recurrent genes provides us strong evidence in understanding the pathophysiology of CP. Limited studies are available that determine the genetic etiologies related to motor type CP because of phenotypic and genotypic heterogeneity. Limited availability of animal model functional studies impedes the understanding of CP pathophysiology. Understanding the association of pro-inflammatory cytokines such as IL-6, IL-1β, and TNF-6 will help in developing important predictive biomarkers for detecting neurodevelopmental disorders. Regulation of Wnt signaling, β-catenin, and GSK-3 pathways suggests that the Wnt/β-catenin signaling process stimulates progenitor cellular proliferation in the development of neural tube, midbrain, and hippocampus during the fetal growth development period. Any impairment of these pathways leads to several neurodevelopmental diseases including CP and autism. A better understanding of all these mechanisms will enable us to identify the risk factors that contribute to CP. Together with the discovery of genetic factors, such as epigenetic and copy number variants, and cytokines regulation, Wnt signaling pathways can provide new opportunities for further detailed analysis and studydriven interventions to improve the lives of children living with CP.

Authors' Contributions

J.U., A.S. and M.N.A. had the idea for this study and designed it and took responsibility for the integrity of the data. J.U. and M.N.A. contributed in the writing of the report. A.S. and M.N.A. contributed to the critical revision of the manuscript. All authors reviewed and approved the final version of the manuscript.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

ORCID iD

Mohd Nazam Ansari D <https://orcid.org/0000-0001-8580-3002>

References

- 1. Bax M, Goldstein M, Rosenbaum P, Leviton A, Paneth N, Dan B, Jacobsson B, Damiano D. Executive Committee for the definition of cerebral palsy. Proposed definition and classification of cerebral palsy, April 2005. *Dev Med Child Neurol* 2005;**47**:571–6
- 2. Ellenberg JH, Nelson KB. The association of cerebral palsy with birth asphyxia: a definitional quagmire. *Dev Med Child Neurol* 2013;**55**:210–6
- 3. Paneth N, Hong T, Korzeniewski S. The descriptive epidemiology of cerebral palsy. *Clin Perinatol* 2006;**33**:251–67
- 4. Hagberg H, Gressens P, Mallard C. Inflammation during fetal and neonatal life: implications for neurologic and neuropsychiatric disease in children and adults. *Ann Neurol* 2012;**71**:444–57
- 5. Badawi N, Felix JF, Kurinczuk JJ, Dixon G, Watson L, Keogh JM, Valentine J, Stanley FJ. Cerebral palsy following term newborn encephalopathy: a population-based study. *Dev Med Child Neurol* 2005;**47**:293–8
- 6. Sweetman DU, Onwuneme C, Watson WR, Murphy JF, Molloy EJ. Perinatal asphyxia and erythropoietin and VEGF: serial serum and cerebrospinal fluid responses. *Neonatology* 2017;**111**:253–9
- 7. Chawanpaiboon S, Vogel JP, Moller AB, Lumbiganon P, Petzold M, Hogan D, Landoulsi S, Jampathong N, Kongwattanakul K, Laopaiboon M, Lewis C, Rattanakanokchai S, Teng DN, Thinkhamrop J, Watananirun K, Zhang J, Zhou W, Gülmezoglu AM. Global, regional, and national estimates of levels of preterm birth in 2014: a systematic review and modelling analysis. *Lancet Glob Health* 2019;**7**:e37–46
- 8. Ball G, Aljabar P, Nongena P, Kennea N, Gonzalez-Cinca N, Falconer S, Chew ATM, Harper N, Wurie J, Rutherford MA, Counsell SJ, Edwards AD. Multimodal image analysis of clinical influences on preterm brain development. *Ann Neurol* 2017;**82**:233–46
- 9. Batalle D, Hughes EJ, Zhang H, Tournier JD, Tusor N, Aljabar P, Wali L, Alexander DC, Hajnal JV, Nosarti C, Edwards AD, Counsell SJ. Early development of structural networks and the impact of prematurity on brain connectivity. *NeuroImage* 2017;**149**:379–92
- 10. Boardman JP, Craven C, Valappil S, Counsell SJ, Dyet LE, Rueckert D, Aljabar P, Rutherford MA, Chew AT, Allsop JM, Cowan F, Edwards AD. A common neonatal image phenotype predicts adverse neurodevelopmental outcome in children born preterm. *NeuroImage* 2010;**52**:409–14
- 11. Galdi P, Blesa M, Stoye DQ, Sullivan G, Lamb GJ, Quigley AJ, Thrippleton MJ, Bastin ME, Boardman JP. Neonatal morphometric similarity mapping for predicting brain age and characterizing neuroanatomic variation associated with preterm birth. *Neuroimage Clin* 2020;**25**:102195
- 12. Kapellou O, Counsell SJ, Kennea N, Dyet L, Saeed N, Stark J, Maalouf E, Duggan P, Ajayi-Obe M, Hajnal J, Allsop JM, Boardman J, Rutherford MA, Cowan F, Edwards AD. Abnormal cortical development after premature birth shown by altered allometric scaling of brain growth. *PLoS Med* 2006;**3**:e265
- 13. Van Naarden Braun K, Doernberg N, Schieve L, Christensen D, Goodman A, Yeargin-Allsopp M. Birth prevalence of cerebral palsy: a population-based study. *Pediatrics* 2016;**137**:1–9
- 14. Costeff H. Estimated frequency of genetic and nongenetic causes of congenital idiopathic cerebral palsy in west Sweden. *Ann Hum Genet* 2004;**68**:515–20
- 15. MacLennan AH, Thompson SC, Gecz J. Cerebral palsy: causes, pathways, and the role of genetic variants. *Am J Obstet Gynecol* 2015;**213**:779–88
- 16. Moreno-De-Luca A, Ledbetter DH, Martin CL. Genetic [corrected] insights into the causes and classification of [corrected] cerebral palsies. *Lancet Neurol* 2012;**11**:283–92
- 17. Fahey MC, Maclennan AH, Kretzschmar D, Gecz J, Kruer MC. The genetic basis of cerebral palsy. *Dev Med Child Neurol* 2017;**59**:462–9
- Derrick M, Drobyshevsky A, Ji X, Tan S. A model of cerebral palsy from fetal hypoxia-ischemia. *Stroke* 2007;**38**:731–5
- 19. Lu B, Zeng F, Xing W, Liang L, Huo J, Tan C, Zhu L, Liu Z. Decreased mitochondrial DNA copy number in children with cerebral palsy quantified by droplet digital PCR. *Clin Chim Acta* 2020;**503**:122–7
- 20. Fryer A, Appleton R, Sweeney MG, Rosenbloom L, Harding AE. Mitochondrial DNA 8993 (NARP) mutation presenting with a heterogeneous phenotype including "cerebral palsy." *Arch Dis Child* 1994; **71**:419–22
- 21. McMichael G, Girirajan S, Moreno-De-Luca A, Gecz J, Shard C, Nguyen LS, Nicholl J, Gibson C, Haan E, Eichler E, Martin CL, MacLennan A. Rare copy number variation in cerebral palsy. *Eur J Hum Genet* 2014;**22**:40–5
- 22. Segel R, Ben-Pazi H, Zeligson S, Fatal-Valevski A, Aran A, Gross-Tsur V, Schneebaum-Sender N, Shmueli D, Lev D, Perlberg S, Blumkin L, Deutsch L, Levy-Lahad E. Copy number variations in cryptogenic cerebral palsy. *Neurology* 2015;**84**:1660–8
- 23. Oskoui M, Gazzellone MJ, Thiruvahindrapuram B, Zarrei M, Andersen J, Wei J, Wang Z, Wintle RF, Marshall CR, Cohn RD, Weksberg R, Stavropoulos DJ, Fehlings D, Shevell MI, Scherer SW. Clinically relevant copy number variations detected in cerebral palsy. *Nat Commun* 2015;**6**:7949
- 24. Mohandas N, Bass-Stringer S, Maksimovic J, Crompton K, Loke YJ, Walstab J, Reid SM, Amor DJ, Reddihough D, Craig JM. Epigenomewide analysis in newborn blood spots from monozygotic twins discordant for cerebral palsy reveals consistent regional differences in DNA methylation. *Clin Epigenetics* 2018;**10**:25
- 25. van Eyk CL, Corbett MA, Gardner A, van Bon BW, Broadbent JL, Harper K, MacLennan AH, Gecz J. Analysis of 182 cerebral palsy transcriptomes points to dysregulation of trophic signalling pathways and overlap with autism. *Transl Psychiatry* 2018;**8**:88
- 26. Jin SC, Lewis SA, Bakhtiari S, Zeng X, Sierant MC, Shetty S, Nordlie SM, Elie A, Corbett MA, Norton BY, van Eyk CL, Haider S, Guida BS, Magee H, Liu J, Pastore S, Vincent JB, Brunstrom-Hernandez J, Papavasileiou A, Fahey MC, Berry JG, Harper K, Zhou C, Zhang J, Li B, Zhao H, Heim J, Webber DL, Frank MSB, Xia L, Xu Y, Zhu D, Zhang B, Sheth AH, Knight JR, Castaldi C, Tikhonova IR, López-Giráldez F, Keren B, Whalen S, Buratti J, Doummar D, Cho M, Retterer K, Millan F, Wang Y, Waugh JL, Rodan L, Cohen JS, Fatemi A, Lin AE, Phillips JP, Feyma T, MacLennan SC, Vaughan S, Crompton KE, Reid SM, Reddihough DS, Shang Q, Gao C, Novak I, Badawi N, Wilson YA, McIntyre SJ, Mane SM, Wang X, Amor DJ, Zarnescu DC, Lu Q, Xing Q, Zhu C, Bilguvar K, Padilla-Lopez S, Lifton RP, Gecz J, MacLennan AH, Kruer MC. Mutations disrupting neuritogenesis genes confer risk for cerebral palsy. *Nat Genet* 2020;**52**:1046–56
- 27. Korzeniewski SJ, Slaughter J, Lenski M, Haak P, Paneth N. The complex aetiology of cerebral palsy. *Nat Rev Neurol* 2018;**14**:528–43
- 28. Gotsch F, Romero R, Kusanovic JP, Mazaki-Tovi S, Pineles BL, Erez O, Espinoza J, Hassan SS. The fetal inflammatory response syndrome. *Clin Obstet Gynecol* 2007;**50**:652–83
- 29. Zareen Z, Strickland T, Eneaney VM, Kelly LA, McDonald D, Sweetman D, Molloy EJ. Cytokine dysregulation persists in childhood post Neonatal Encephalopathy. *BMC Neurol* 2020;**20**:115
- 30. Huang HC, Wang CL, Huang LT, Chuang H, Liu CA, Hsu TY, Ou CY, Yang KD. Association of cord blood cytokines with prematurity and cerebral palsy. *Early Hum Dev* 2004;**77**:29–36
- 31. Keelan JA, Marvin KW, Sato TA, Coleman M, McCowan LM, Mitchell MD. Cytokine abundance in placental tissues: evidence of inflammatory activation in gestational membranes with term and preterm parturition. *Am J Obstet Gynecol* 1999;**181**:1530–6
- 32. Vidak HK, Ivković TC, Jokić M, Spaventi R, Kapitanović S. The association between pro-inflammatory cytokine polymorphisms and cerebral palsy in very preterm infants. *Cytokine* 2012;**58**:57–64

33. Kumral A, Baskin H, Yesilirmak DC, Ergur BU, Aykan S, Genc S, Genc K, Yilmaz O, Tugyan K, Giray O, Duman N, Ozkan H. Erythropoietin attenuates lipopolysaccharide-induced white matter injury in the neonatal rat brain. *Neonatology* 2007;**92**:269–78

- 34. Beloosesky R, Weiner Z, Ginsberg Y, Ross MG. Maternal N-acetylcysteine (NAC) protects the rat fetal brain from inflammatory cytokine responses to lipopolysaccharide (LPS). *J Matern Fetal Neonatal Med* 2012;**25**:1324–8
- 35. Massaro G, Scaravilli G, Simeone S, Capuano S, Pastore E, Forte A, Parisi P, Ferraiolo A, Costanzo A, Balbi C. Interleukin-6 and *Mycoplasma hominis* as markers of preterm birth and related brain damage: our experience. *J Matern Fetal Neonatal Med* 2009;**22**:1063–7
- 36. Falahati S, Breu M, Waickman AT, Phillips AW, Arauz EJ, Snyder S, Porambo M, Goeral K, Comi AM, Wilson MA, Johnston MV, Fatemi A. Ischemia-induced neuroinflammation is associated with disrupted development of oligodendrocyte progenitors in a model of periventricular leukomalacia. *Dev Neurosci* 2013;**35**:182–96
- 37. Yoneda S, Shiozaki A, Ito M, Yoneda N, Inada K, Yonezawa R, Kigawa M, Saito S. Accurate Prediction of the Stage of Histological Chorioamnionitis before Delivery by Amniotic Fluid IL-8 Level. *Am J Reprod Immunol* 2015;**73**:568–76
- 38. Strackx E, Sparnaaij MA, Vlassaks E, Jellema R, Kuypers E, Vles JS, Kramer BW, Gavilanes AW. Lipopolysaccharide-induced chorioamnionitis causes acute inflammatory changes in the ovine central nervous system. *CNS Neurol Disord Drug Targets* 2015;**14**:77–84
- 39. Rodgers JM, Robinson AP, Rosler ES, Lariosa Willingham K, Persons RE, Dugas JC, Miller SD. IL-17 A activates ERK 1/2 and enhances differentiation of oligodendrocyte progenitor cells. *Glia* 2015;**63**:768–79
- 40. Yan X, Sun M, Gibb W. Localization of nuclear factor-kappa B (NF kappa B) and inhibitory factor-kappa B (I kappa B) in human fetal membranes and decidua at term and preterm delivery. *Placenta* 2002;**23**:288–93
- 41. Lappas M, Permezel M, Georgiou HM, Rice GE. Nuclear factor kappa B regulation of proinflammatory cytokines in human gestational tissues in vitro. *Biol Reprod* 2002;**67**:668–73
- 42. Hansen-Pupp I, Hallin AL, Hellström-Westas L, Cilio C, Berg AC, Stjernqvist K, Fellman V, Ley D. Inflammation at birth is associated with subnormal development in very preterm infants. *Pediatr Res* 2008;**64**:183–8
- 43. Moylan JS, Smith JD, Wolf Horrell EM, McLean JB, Deevska GM, Bonnell MR, Nikolova-Karakashian MN, Reid MB. Neutral sphingomyelinase-3 mediates TNF-stimulated oxidant activity in skeletal muscle. *Redox Biol* 2014;**2**:910–20
- 44. Yawno T, Schuilwerve J, Moss TJ, Vosdoganes P, Westover AJ, Afandi E, Jenkin G, Wallace EM, Miller SL. Human amnion epithelial cells reduce fetal brain injury in response to intrauterine inflammation. *Dev Neurosci* 2013;**35**:272–82
- 45. Lu S, Xiao X, Cheng M. Matrine inhibits IL-1β-induced expression of matrix metalloproteinases by suppressing the activation of MAPK and NF-κB in human chondrocytes in vitro. *Int J Clin Exp Pathol* 2015; **8**:4764–72
- 46. Silva ML, Ribeiro AP, Silva GA, Sanchez IX, Renzo R, Uscategui R, Lima TB, Aldrovani M, Laus JL. Expressions of matrix metalloproteinases-1 and -9 and opioid growth factor in rabbit cornea after lamellar keratectomy and treatment with 1% nalbuphine. *Arq Bras Oftalmol* 2015;**78**:141–5
- 47. Hansen-Pupp I, Harling S, Berg AC, Cilio C, Hellström-Westas L, Ley D. Circulating interferon-gamma and white matter brain damage in preterm infants. *Pediatr Res* 2005;**58**:946–52
- 48. Di H, He Q, Liao Y, Kalionis B, Tai X. The role of inflammatory cytokines in the pathogenesis of cerebral palsy. *Gynecol Obstet* 2016;**6**:360
- 49. Wen F, Tao W, Yao H, Sun Y. The influence of erythropoietin and proinflammatory cytokines in the development of cerebral palsy. *Vasc Dis Prev* 2008;**5**:29–32
- 50. Harrigan MR, Ennis SR, Masada T, Keep RF. Intraventricular infusion of vascular endothelial growth factor promotes cerebral angiogenesis with minimal brain edema. *Neurosurgery* 2002;**50**:589–98
- 51. Petersen CP, Reddien PW. Wnt signaling and the polarity of the primary body axis. *Cell* 2009;**139**:1056–68
- 52. Kim WY, Snider WD. Functions of GSK-3 signaling in development of the nervous system. *Front Mol Neurosci* 2011;**4**:44
- 53. Yeung TM, Chia LA, Kosinski CM, Kuo CJ. Regulation of self-renewal and differentiation by the intestinal stem cell niche. *Cell Mol Life Sci* 2011;**68**:2513–23
- 54. Budnik V, Salinas PC. Wnt signaling during synaptic development and plasticity. *Curr Opin Neurobiol* 2011;**21**:151–9
- 55. Valvezan AJ, Klein PS. GSK-3 and Wnt signaling in neurogenesis and bipolar disorder. *Front Mol Neurosci* 2012;**5**:1–13
- 56. Kim YT, Hur EM, Snider WD, Zhou FQ. Role of GSK3 signaling in neuronal morphogenesis. *Front Mol Neurosci* 2011;**4**:48
- 57. Ng SS, Mahmoudi T, Danenberg E, Bejaoui I, de Lau W, Korswagen HC, Schutte M, Clevers H. Phosphatidylinositol 3-kinase signaling does not activate the wnt cascade. *J Biol Chem* 2009;**284**:35308–13
- 58. McManus EJ, Sakamoto K, Armit LJ, Ronaldson L, Shpiro N, Marquez R, Alessi DR. Role that phosphorylation of GSK3 plays in insulin and Wnt signalling defined by knockin analysis. *EMBO J* 2005;**24**:1571–83
- 59. Ciani L, Salinas PC. WNTs in the vertebrate nervous system: from patterning to neuronal connectivity. *Nat Rev Neurosci* 2005;**6**:351–62
- 60. Mudher A, Shepherd D, Newman TA, Mildren P, Jukes JP, Squire A, Mears A, Drummond JA, Berg S, MacKay D, Asuni AA, Bhat R, Lovestone S. GSK-3beta inhibition reverses axonal transport defects and behavioural phenotypes in *Drosophila. Mol Psychiatry* 2004;**9**:522–30
- 61. Kapsimali M, Caneparo L, Houart C, Wilson SW. Inhibition of Wnt/ Axin/beta-catenin pathway activity promotes ventral CNS midline tissue to adopt hypothalamic rather than floorplate identity. *Development* 2004;**131**:5923–33
- 62. Muroyama Y, Fujihara M, Ikeya M, Kondoh H, Takada S. Wnt signaling plays an essential role in neuronal specification of the dorsal spinal cord. *Genes Dev* 2002;**16**:548–53
- 63. Megason SG, McMahon AP. A mitogen gradient of dorsal midline Wnts organizes growth in the CNS. *Development* 2002;**129**:2087–98
- 64. Panhuysen M, Vogt Weisenhorn DM, Blanquet V, Brodski C, Heinzmann U, Beisker W, Wurst W. Effects of Wnt1 signaling on proliferation in the developing mid-/hindbrain region. *Mol Cell Neurosci* 2004;**26**:101–11
- 65. Zechner D, Fujita Y, Hülsken J, Müller T, Walther I, Taketo MM, Crenshaw EB, 3rd Birchmeier W, Birchmeier C. beta-Catenin signals regulate cell growth and the balance between progenitor cell expansion and differentiation in the nervous system. *Dev Biol* 2003;**258**:406–18
- 66. Lee SM, Tole S, Grove E, McMahon AP. A local Wnt-3a signal is required for development of the mammalian hippocampus. *Development* 2000;**127**:457–67
- 67. Machon O, van den Bout CJ, Backman M, Kemler R, Krauss S. Role of beta-catenin in the developing cortical and hippocampal neuroepithelium. *Neuroscience* 2003;**122**:129–43
- 68. Purro SA, Ciani L, Hoyos-Flight M, Stamatakou E, Siomou E, Salinas PC. Wnt regulates axon behavior through changes in microtubule growth directionality: a new role for adenomatous polyposis coli. *J Neurosci* 2008;**28**:8644–54
- 69. Lucas FR, Salinas PC. WNT-7a induces axonal remodeling and increases synapsin I levels in cerebellar neurons. *Dev Biol* 1997;**192**:31–44
- 70. Hall AC, Lucas FR, Salinas PC. Axonal remodeling and synaptic differentiation in the cerebellum is regulated by WNT-7a signaling. *Cell* 2000;**100**:525–35
- 71. Dill J, Wang H, Zhou F, Li S. Inactivation of glycogen synthase kinase 3 promotes axonal growth and recovery in the CNS. *J Neurosci* 2008;**28**:8914–28
- 72. Wolf AM, Lyuksyutova AI, Fenstermaker AG, Shafer B, Lo CG, Zou Y. Phosphatidylinositol-3-kinase-atypical protein kinase C signaling is required for Wnt attraction and anterior-posterior axon guidance. *J Neurosci* 2008;**28**:3456–67
- 73. Ciani L, Boyle KA, Dickins E, Sahores M, Anane D, Lopes DM, Gibb AJ, Salinas PC. Wnt7a signaling promotes dendritic spine growth and synaptic strength through Ca²⁺/calmodulin-dependent protein kinase II. *Proc Natl Acad Sci USA* 2011;**108**:10732–7
- 74. Penagini F, Mameli C, Fabiano V, Brunetti D, Dilillo D, Zuccotti GV. Dietary intakes and nutritional issues in neurologically impaired children. *Nutrients* 2015;**7**:9400–15
- 75. Fung EB, Samson-Fang MD, Stallings MD, Conaway M, Liptak MD, Henderson RC, Worley G, O'Donnell M, Calvert R, Rosenbaum P, Chumlea W, Stevenson RD. Feeding dysfunction is associated with poor growth and health status in children with cerebral palsy. *J Am Diet Assoc* 2002;**102**:361–73
- 76. Ferluga ED, Sathe NA, Krishnaswami S, Mcpheeters ML. Surgical intervention for feeding and nutrition difficulties in cerebral palsy: a systematic review. *Dev Med Child Neurol* 2014;**56**:31–43
- 77. Kim HJ, Choi HN, Yim JE. Food habits, dietary intake, and body composition in children with cerebral palsy. *Clin Nutr Res* 2018;**7**:266–75
- 78. Hariprasad PG, Elizabeth KE, Valamparampil MJ, Kalpana D, Anish TS. Multiple nutritional deficiencies in cerebral palsy compounding physical and functional impairments. *Indian J Palliat Care* 2017;**23**:387–92