Minireview

Dysregulation of brain dopamine systems in major depressive disorder

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Impact statement

MDD is produced by complex and heterogeneous disruptions in brain architecture, neuroplasticity, and function, with genetic, epigenetic, and environmental contributions. In addition to other biogenic amines, the symptoms and pathophysiology of depression indicate significant involvement of dysfunction in dopamine and dopaminergic circuits. New studies are providing a much more precise and nuanced view of how depression may originate from neurodevelopmental maladaptations in dopamine-modulated processes, and suggest new targets for therapeutic intervention.

Abstract

Major depressive disorder (MDD or depression) is a debilitating neuropsychiatric syndrome with genetic, epigenetic, and environmental contributions. Depression is one of the largest contributors to chronic disease burden; it affects more than one in six individuals in the United States. A wide array of cellular and molecular modifications distributed across a variety of neuronal processes and circuits underlie the pathophysiology of depression—no established mechanism can explain all aspects of the disease. MDD suffers from a vast treatment gap worldwide, and large numbers of individuals who require treatment do not receive adequate care. This mini-review focuses on dysregulation of brain dopamine (DA) systems in the pathophysiology of MDD and describing new cellular targets for potential medication development focused on DA-modulated micro-circuits. We also explore how neurodevelopmental factors may modify risk for later emergence of MDD, possibly through

dopaminergic substrates in the brain.

Keywords: Depression, major depressive disorder, receptor, neurodevelopment, anhedonia, mood

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Major depressive disorder

Major depressive disorder (MDD) is one of the world's greatest public health concerns due to its prevalence, impact, and complexity. MDD, otherwise known as depression or clinical depression, is defined as a prolonged period of time of a person experiencing a depressed mood, loss of interest, or pleasure.¹ MDD affects at least one in six people in the US population,² and a 2007 World Health Organization study of over 200,000 adults demonstrated that MDD results in the greatest overall reduction in health when compared with other debilitating and chronic diseases, including diabetes and arthritis. 3 MDD encompasses a variety of symptoms related to mood, cognition, and hedonic systems that implicate multiple central nervous system (CNS) circuits and neural functions.^{4,5} These symptoms include anxiety, fatigue, anhedonia, changes in sleep and activity, and even suicidality. It is difficult to ascertain which CNS changes contribute to the underlying pathophysiology of MDD, rather than function as compensatory neuroplastic responses induced by the CNS to

ameliorate disease processes. Early studies of the neurobiology of MDD focused largely on the neurotransmitters norepinephrine and serotonin, due to the antidepressant properties of pro-noradrenergic and pro-serotonergic agents, including the selective serotonin reuptake inhibitors (SSRIs). However, serotonergic dysfunction alone is insufficient to fully describe MDD and its treatment (for excellent reviews of this topics, please see references^{6,7}). Current antidepressants are associated with only moderate efficacy, delayed therapeutic effects, and significant side effects. Approximately 30% of patients do not remit from MDD, even after several treatment attempts. Moreover, newly developed medications show a high failure rate in clinical trials, and a significant proportion of MDD patients are "treatment-resistant".⁸

Recent studies on MDD have explored several intriguing and important avenues in this regard. First, genetic studies have continued to identify multiple risk alleles and diseasemodifying genes across a wide variety of neurochemical and cellular function families.⁹ Important environmental,

behavioral, developmental, and epigenetic factors continue to be identified, 10^{-12} and a more detailed understanding of altered brain architecture is now evident.¹³ There is still a pressing need for fast-acting, specific, and efficacious therapies—both pharmacological and behavioral.

Early studies of the monoamine hypothesis of MDD also considered dopamine (DA), but interest in DA soon focused on movement disorders and drug use and abuse. Recently, the crucial role of anhedonia as an endophenotype of MDD has been re-emphasized, and indeed, this is a cardinal symptom of MDD. Anhedonia is unlikely to be a specific symptom of MDD; for example, it is also prevalent in disorders historically linked conceptually to DA dysfunction, such as schizophrenia and Parkinson's disease.^{14,15} This also encompasses the complex rewardrelated deficits observed in MDD such as disruption of decisional anhedonia, which refers to the anticipation, motivation, and decision-making process involved in obtaining a reward.16,17 Importantly, anhedonia is one the hallmark symptoms of MDD aside from depressed mood, and was shown nearly 50 years ago to be a sign of poor antidepressant response.¹⁸ No treatment has been efficient in treating anhedonia; in this regard SSRIs have been shown to be ineffective for positive affect deficits.¹⁹ In this mini-review, therefore, we focus on dysregulation of brain DA systems in the pathophysiology of MDD and propose new cellular targets for potential medication development focused on DA-modulated micro-circuits and novel cellular targets. We also explore how neurodevelopmental factors may modify risk for later emergence of MDD, possibly through DAergic substrates in the developing brain.

Dopamine and dopamine receptors

DA is a catecholamine neurotransmitter and neuromodulator that primarily acts through binding to high-affinity receptors. DA receptors (D1–D5) are G-protein coupled receptors with relatively slow effects on synaptic signaling through second messenger systems. DAergic systems modulate a wide variety of neural functions and behaviors, including motor control, motivation, reward, cognition, and maternal and reproductive behaviors. DA receptors are divided into two primary families: the D1-like and the D2-like receptors. The D1-like receptor family includes the D1 and D5 receptors, which are encoded by the DRD1 and DRD5 genes, respectively. The D2-like receptor family includes the D2, D3, and D4 receptors, which are similarly encoded by DRD1, DRD2, and DRD4, respectively. Each subtype of DA receptor has a unique regional and cellular pattern of expression, developmental ontogeny, regulatory properties, and functional roles. The two families (D1-like and D2-like) are easily distinguished pharmacologically, but development of truly specific agonists and antagonists of each molecular receptor subtype has been challenging for the field. D1 and D2 receptors are the most abundantly expressed receptors, and are rarely co-expressed in the same neurons, at least in adult animals.²⁰ D1-like receptor activation can contribute to either excitation or inhibition, dependent on cell type; D2-like receptor activation is usually inhibitory.^{21–23} DA signaling is typically terminated by

re-uptake of DA through a high-affinity presynaptic transporter (DAT).

The D1-like receptors typically stimulate adenylate cyclase to increase the intracellular concentration of the second messenger cyclic adenosine monophosphate downstream of activation of the adenylate cyclase stimulatory G proteins $G_{\alpha s/\alpha o l f}$ this, in turn, stimulates the activity of protein kinase A.^{24,25} D1 receptors are mostly found in the caudate and putamen (striatum), substantia nigra pars reticulata, nucleus accumbens (NAc), olfactory bulb, amygdala, and frontal cortex, with D5 receptors particularly expressed in the cerebral cortex.^{26,27} On the other hand, D2, D3, and D4 receptors all couple to $G_{\alpha i/\alpha o}$ -proteins to primarily inhibit adenylate cyclase.²⁸ This group of receptors is mainly expressed in the striatum, cerebral cortex, hippocampus, NAc, substantia nigra pars compacta, and pituitary gland.27,29,30 D2-like receptors can also modulate the Akt-GSK3 signaling pathway which in turn regulates proliferation, differentiation, and gene transcription.³¹ MAPK signaling is also modulated by both D1- and D2 like receptors, and this pathway significantly contributes to DA-mediated modulation of cell death, developmental patterning, and synaptic plasticity.³²

There are several major DAergic pathways in the brain.33,34 The main brain regions containing DAcontaining cell bodies are in the midbrain and consist of the substantia nigra pars compacta (SN) and the ventral tegmental area (VTA). Axons arising from the DA cells in the SN pars compacta form the nigro-striatal tract and provide DA innervation to the dorsal striatum, arranged in a topographical manner. The striatum is part of the basal ganglia, a group of forebrain structures involved in motor control, motivation, and cognition. Degeneration of the nigrostriatal DA neurons is the main pathology of the movement disorder Parkinson's disease, but Parkinson's disease is also characterized by dysfunctions in affect, mood, and reward. The mesolimbic and mesofrontocortical DA systems arise from the medially localized VTA. The mesolimbic system provides DA innervation to subcortical regions including the NAc, septum, olfactory tubercle, hippocampus, and amygdala. The NAc (sometimes also referred to as the ventral striatum) is an important interface for functional output related to motivation and motor systems. The NAc consists of two subregions containing largely GABAergic medium spiny neurons, the core and the shell, and it is a vital component of brain reward systems. The mesofrontocortical system provides dopaminergic afferents to specific cerebral cortical areas, including the orbitofrontal and medial prefrontal cortex (PFC). Disruption of DA neurotransmission within the mesofrontocortical system has been associated with mental health conditions, including schizophrenia, bipolar depression, substance use disorders, and MDD. The PFC is involved in complex cognitive functions including motor planning, attention, and behavioral inhibition and is highly stresssensitive (see below). The PFC sends glutamatergic projections to forebrain structures including the striatum, midbrain, and hippocampus. Lastly, the tuberoinfundibular DA system is an important hypothalamic pathway,

which connects the hypothalamus and pituitary gland to control prolactin secretion in the anterior pituitary.

Dopamine, stress, and MDD

Roles for the dopamine system in modulating stress and depression-related circuits have been posited for some time (for reviews, see references $35-38$). Mesolimbic and mesocortical DA neurons are activated by acute behavioral stressors, as well as by other behaviorally salient stimuli. $39-42$ Mesocortical DA neurons are especially sensitive to acute stressful stimuli such that relatively mild stressors induce marked activation of these neurons and of DA neurotransmission in the PFC. Functional heterogeneity is observed within the mesolimbic projections to the NAc in that behavioral stressors activate DAergic activity to a greater extent in the NAc shell than in the NAc core or dorsal striatum.⁴³ Studies of conditioned fear also implicate mesolimbic DA pathways as a key controller of fear-related learning, and DA-recipient cell targets in the NAc have been directly implicated in animal models of depression.^{36,44} Acute exposure to stress has been shown to increase DA synthesis activity in terminal regions and the rank order of this effect again reflects differential responses across DA circuits (PFC greater than NAc; NAc greater than dorsal striatum). Stress also serves as a crucial potential of drug self-administration and relapse. Collectively, these data demonstrate that region and cell-specific increases in DA neurotransmission and downstream signaling activation occur during exposure to acute stress. Studies in humans reveal that blocking or decreasing DA using pharmacological interventions resulted in induction and deepening of depression.³⁵ Conversely, antidepressants increase brain DAergic mechanisms 45 and D2-like receptor antagonists are often efficacious successful adjuvants to SSRIs in the treatment of MDD.⁴⁶ For example, lurasidone and aripiprazole/brexpiprazole are new agents with DA receptor modulating effects that are both effective in treating depressive symptoms.46–49

Rodent models of MDD also implicate and replicate DAergic contributions to MDD-induced cellular and regional pathologies and maladaptations (Table 1; see also the following excellent reviews focused on this topic $50-52$). Anhedonia, again, takes center stage, and this can be assessed through a variety of assays with distinct strengths and limitations, ranging from simple sucrose consumption assays to complex operant models of decisional anhedonia.5,17,53–58 Because MDD has a such a complex etiology across individuals, animal models similarly utilize a number of induction agents, including acute and chronic stress exposure, early life stress and/or maternal neglect, exogenous administration of glucocorticoids, chronic inflammatory conditions, and genetic manipulations $59-61$ (Table 1). Chronic mild unpredictable stress, social defeat stress, and resident-intruder chronic social stress appear to have particularly strong validity and alter a wide variety of neural circuits and behaviors, including CNS DA systems and their targets.⁶²⁻⁶⁹ However, others have argued that laboratory rodents do not encounter the adaptive evolutionary social pressures required for the development of depression, and thus all rodent models of MDD may be irrevocably flawed.⁷⁰

Several key recent studies have brought a focus back to dopaminergic systems, and DA D1 receptors specifically, in considering the underlying pathophysiology of mood disorders. Optogenetic initiation of phasic firing patterns in VTA dopaminergic neurons projecting to the medial PFC induced increased susceptibility to social defeat stress.⁶⁵ Another group instead optogenetically inhibited VTA neurons in awake, behaving mice and observed that this instantly reduced struggling in the tail suspension test and induced anhedonia in a sucrose preference test. 71 Within the cerebral cortex itself, overexpression of D1 receptors only slightly increased sucrose preference, but termination of the overexpression (to normal D1 levels) produced profound depression-like behaviors across several paradigms.⁷² Several additional studies also implicate dopaminoceptive circuits within the PFC, hippocampus, and amygdala in altering responses to challenges and stressors.^{73–75} Beyond MDD, alterations in dopaminergic circadian activity may contribute to rapid mood-cycling in bipolar disorder.⁷⁶ A newly identified population of D1 receptor-expressing neurons within a subregion of the medial amygdala determines approach-avoidance conflict in the face of threatening stimuli.⁷⁷ A previously neglected population of DA neurons in the dorsal raphe nucleus, the brain region usually associated with 5-HT neurons and the actions of many antidepressant drugs, has recently been demonstrated to functionally contribute to aspects of mood regulation.⁷⁸ A newly identified DA-modulated circuit involving D1 receptor-containing neurons in the interpeduncular nucleus contributes vitally to the regulation of anxiety behaviors.⁷⁹ In this newly identified circuit, VTA projections to the interpeduncular nucleus activate D1 receptors to modulate a specific subtype of GABAergic neuron, which then differentially excite or inhibit two additional specific subpopulations of neurons to drive anxiety-related behavior. Perhaps most strikingly, D1 receptor-mediated changes in the structure and function of PFC neurons can suppress stress susceptibility.⁸⁰ Emerging data from our group indicate that cerebral cortical interneuron deletion of D1 receptors produces mice who have strong antidepressant and stress resilient phenotypes at baseline (Delva and Stanwood, unpublished observations). These data suggest that cell-specific neuropharmacology strategies are needed, location- and signaling-biased ligands may be particularly useful.⁸¹ For example, a ligand that blocks D1 receptors expressed on cortical interneurons, but not D1 receptors expressed by excitatory neurons, striatal medium-spiny neurons, or interpeduncular nucleus neurons would represent a new and exciting mechanism to treat MDD. These studies are also summarized in Table 2.

Individuals with MDD also exhibit changes in markers of DAergic circuits and neurochemical regulation. For example, in vivo availability of DAT, as studied with a PET tracer, is reduced both in the putamen and VTA, bilaterally.⁸² Reduced DAT levels after stressor exposure have been observed in animal studies as well, and this effect is typically interpreted as reflecting compensatory DAT

Table 1. Animal models, especially rodent models, are a crucial step in understanding the pathophysiology and treatment of MDD and other depression-related disorders, but also have limitations.^a

Table 1. Continued.

^aAlso see the following excellent reviews focused on this topic.⁵⁰⁻⁵²

Table 2. Summary table of recent key animal studies establishing new specific roles for dopaminergic mechanisms in mood-related circuits and phenotypes.

downregulation following blunted DA neurotransmission. Postmortem Western blot analyses confirm the reduction in DAT in the putamen and VTA, but not in the caudate or NAc, and further demonstrate reduced tyrosine hydroxylase, the rate limiting enzyme in DA synthesis. 82 Moreover, the number of depressive episodes and specific symptom presentation within individuals was predictive of the decreases in DAT availability. Individuals with MDD who reported feeling trapped in perceived inescapable circumstances showed the lowest DAT availability in the VTA, suggesting that DAergic impairment might be more prevalent in a subset of people with severe and specific MDD symptoms. Replication of these findings are needed. Importantly, this study focused on unmedicated individuals with MDD and minimal comorbidity with other conditions; these issues have made it difficult to find consistency of findings in human MDD patients. Additional studies have documented reduced reward-related neural fMRI signals in the caudate and $NAc₁⁸³$ and alterations in both striatal and cerebral cortical responses to positive mood states.⁸⁴ Notably then, significant heterogeneity has been observed among studies, with some changes localized to DA cell body-containing regions, and other to terminals and/or dopaminoceptive targets. In addition, some changes localize more strongly to different striatal subdivisions; it is unclear whether this reflects differences in study subjects, in circuit-related neuroadaptations, or in the specific brain mechanisms interrogated by different measurements. Of additional interest, expression of the gene CHRNA7, which encodes a subunit of the nicotinic acetylcholine receptor and regulates cognition through interneuron modulation of DA signaling, is increased in the PFC of patients with MDD.⁸⁵ Conversely, protection from MDD following stress can be conferred through resiliency mechanisms. Recent studies of the neurobiology of resiliency indicate that the function of the mesolimbic DA system contributes to adaptive behavioral responses to stress.⁸⁶

Neurodevelopmental origins of MDD and DA dysregulation during brain development

MDD and other neuropsychiatric disorders are increasingly being viewed from a neurodevelopmental perspective, with an understanding that early alterations in neuronal migration, differentiation, and circuit formation provides an altered CNS substrate on which additional deleterious factors in childhood and adolescence can further dysregulate to produce disease.^{87,88} Recent studies in animal models and humans strongly implicate cortical GABAergic neuronal development in mood regulation and the pathophysiology of major depression.⁸⁹⁻⁹³ But as is the case with most neuronal substrates, GABAergic interneurons in the cortex are embryonically, structurally, and functionally diverse, regulating a very delicate balance across cortical networks.⁹⁴⁻⁹⁷ Of note, PFC interneurons receive synaptic input from dopaminergic axons, and express DA D1 and D2 receptors. $90,98-101$

In addition to being a neurotransmitter in the adult CNS, DA has important modulatory roles during pre- and postnatal development.¹⁰² Conversely, altering neural circuits and behavioral experiences during neurodevelopment can dramatically alter the stabilizing and adaptive influences of central DA systems in the adult. For example, pre- or early postnatal exposure to stressors, drugs of abuse, or environmental toxins enduringly alter the neuroanatomical organization, cellular responsiveness, and adaptive plasticity of both DA neurons themselves and their forebrain targets.103,104 Direct neurodevelopmental roles for genetic and environmental modulation of DA receptors and signaling components have been described by multiple laboratories and in multiple mammalian species. $105-113$ The quality of parental care has also been observed to alter these mechanisms in multiple animal species/models, as well as in people. For example, repeated epochs of maternal separation during early postnatal development decrease DAT activity and increase systemic and CNS DA responses to subsequent stressors.¹¹⁴⁻¹¹⁶ DA systems are also extremely sensitive to genetic, environmental, and drug-induced modifications during adolescent development.^{117,118} Thus, elucidation of these risk factors, their varied but specific effects on neurodevelopmental trajectory, and the development of therapeutic methods to stabilize brain maturation and adaptive plasticity even in the face of such risk factors, are all vital areas of future study in the pathophysiology of MDD and its associated phenotypes.

Delayed DA maturation, connectivity, and plasticity during adolescence may be involved in amplifying the risk to developing mental disorders, particularly in the PFC.^{38,119,120} Consequently, environmental influences may disturb adolescent prefrontal dopamine development which eventually either worsen or render individuals to become resistant to certain psychiatric conditions, including MDD and schizophrenia.

Summary

MDD is produced by complex and heterogeneous disruptions in neurobiology with genetic, epigenetic, and environmental contributions. CNS biogenic amines, including DA, have important stabilizing, integrative, and plasticityrelated influences on brain circuits. Biogenic amine disruptions are certainly a component of the pathophysiology of MDD, although it remains poorly understood which of these are causal and which may be consequence. Based on emerging studies from animal models and clinical studies, we propose that the brain DA system may be crucially involved in underlying both neuropathological processes and in promoting adaptive mechanisms contributing to resilience, particularly during sensitive periods of development. New technologies and understanding are allowing researchers to frame much more specific hypotheses than ever before. The creation of neuronal cell-type specific pharmacological ligands, coupled with a modern understanding of underlying sub-circuits and neuroplastic mechanisms, has the ability to allow us to take the management of this disabling disorder to the next level. Ultimately, the heterogenous spectrum of clinical MDD requires a diversity of therapeutic options and specific modulation of DA sub-circuits within the CNS will add measurably to our pharmacological treatment repertoire.

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All authors participated in the writing and review of the manuscript.

DECLARATION OF CONFLICTING INTERESTS

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REFERENCES

- 1. American Psychiatric Association DSMTF. In: American Psychiatric A, American Psychiatric Association DSMTF (eds) Diagnostic and statistical manual of mental disorders: DSM-5. Arlington, VA: American Psychiatric Association, 2013
- 2. Kessler RC, Berglund P, Demler O, Jin R, Koretz D, Merikangas KR, Rush AJ, Walters EE, Wang PS. The epidemiology of major depressive disorder: results from the National Comorbidity Survey Replication (NCS-R). JAMA 2003;289:3095–105
- 3. Moussavi S, Chatterji S, Verdes E, Tandon A, Patel V, Ustun B. Depression, chronic diseases, and decrements in health: results from the World Health Surveys. Lancet 2007;370:851-8
- 4. Belujon P, Grace AA. Dopamine system dysregulation in major depressive disorders. Int J Neuropsychopharmacol 2017;20:1036–46
- 5. Heshmati M, Russo SJ. Anhedonia and the brain reward circuitry in depression. Curr Behav Neurosci Rep 2015;2:146–53
- 6. Liu B, Liu J, Wang M, Zhang Y, Li L. From serotonin to neuroplasticity: evolvement of theories for major depressive disorder. Front Cell Neurosci 2017;11:305
- 7. Kupfer DJ, Frank E, Phillips ML. Major depressive disorder: new clinical, neurobiological, and treatment perspectives. Lancet 2012;379:1045–55
- 8. Akil H, Gordon J, Hen R, Javitch J, Mayberg H, McEwen B, Meaney MJ, Nestler EJ. Treatment resistant depression: a multi-scale, systems biology approach. Neurosci Biobehav Rev 2018;84:272–88
- 9. Wray NR, Ripke S, Mattheisen M, Trzaskowski M, Byrne EM, Abdellaoui A, Adams MJ, Agerbo E, Air TM, Andlauer TMF, Bacanu SA, Bækvad-Hansen M, Beekman AFT, Bigdeli TB, Binder EB, Blackwood DRH, Bryois J, Buttenschøn HN, Bybjerg-Grauholm J, Cai N, Castelao E, Christensen JH, Clarke TK, Coleman JIR, Colodro-Conde L, Couvy-Duchesne B, Craddock N, Crawford GE, Crowley CA, Dashti HS, Davies G, Deary IJ, Degenhardt F, Derks EM, Direk N, Dolan CV, Dunn EC, Eley TC, Eriksson N, Escott-Price V, Kiadeh FHF, Finucane HK, Forstner AJ, Frank J, Gaspar HA, Gill M, Giusti-Rodríguez P, Goes FS, Gordon SD, Grove J, Hall LS, Hannon E, Hansen CS, Hansen TF, Herms S, Hickie IB, Hoffmann P, Homuth G, Horn C, Hottenga JJ, Hougaard DM, Hu M, Hyde CL, Ising M, Jansen R, Jin F, Jorgenson E, Knowles JA, Kohane IS, Kraft J, Kretzschmar WW, Krogh J, Kutalik Z, Lane JM, Li Y, Li Y, Lind PA, Liu X, Lu L, MacIntyre DJ, MacKinnon DF, Maier RM, Maier W, Marchini J, Mbarek H, McGrath P, McGuffin P, Medland SE, Mehta D, Middeldorp CM, Mihailov E, Milaneschi Y, Milani L, Mill J, Mondimore FM, Montgomery GW, Mostafavi S, Mullins N, Nauck M, Ng B, Nivard MG, Nyholt DR, O'Reilly PF, Oskarsson H,

Owen MJ, Painter JN, Pedersen CB, Pedersen MG, Peterson RE, Pettersson E, Peyrot WJ, Pistis G, Posthuma D, Purcell SM, Quiroz JA, Qvist P, Rice JP, Riley BP, Rivera M, Saeed Mirza S, Saxena R, Schoevers R, Schulte EC, Shen L, Shi J, Shyn SI, Sigurdsson E, Sinnamon GBC, Smit JH, Smith DJ, Stefansson H, Steinberg S, Stockmeier CA, Streit F, Strohmaier J, Tansey KE, Teismann H, Teumer A, Thompson W, Thomson PA, Thorgeirsson TE, Tian C, Traylor M, Treutlein J, Trubetskoy V, Uitterlinden AG, Umbricht D, Van der Auwera S, van Hemert AM, Viktorin A, Visscher PM, Wang Y, Webb BT, Weinsheimer SM, Wellmann J, Willemsen G, Witt SH, Wu Y, Xi HS, Yang J, Zhang F, Arolt V, Baune BT, Berger K, Boomsma DI, Cichon S, Dannlowski U, de Geus ECJ, DePaulo JR, Domenici E, Domschke K, Esko T, Grabe HJ, Hamilton SP, Hayward C, Heath AC, Hinds DA, Kendler KS, Kloiber S, Lewis G, Li QS, Lucae S, Madden PFA, Magnusson PK, Martin NG, McIntosh AM, Metspalu A, Mors O, Mortensen PB, Müller-Myhsok B, Nordentoft M, Nöthen MM, O'Donovan MC, Paciga SA, Pedersen NL, Penninx B, Perlis RH, Porteous DJ, Potash JB, Preisig M, Rietschel M, Schaefer C, Schulze TG, Smoller JW, Stefansson K, Tiemeier H, Uher R, Völzke H, Weissman MM, Werge T, Winslow AR, Lewis CM, Levinson DF, Breen G, Børglum AD, Sullivan PF. Genome-wide association analyses identify 44 risk variants and refine the genetic architecture of major depression. Nat Genet 2018;50:668–81

- 10. Uchida S, Yamagata H, Seki T, Watanabe Y. Epigenetic mechanisms of major depression: targeting neuronal plasticity. Psychiatry Clin Neurosci 2018;72:212–27
- 11. Hoffmann A, Sportelli V, Ziller M, Spengler D. Epigenomics of major depressive disorders and schizophrenia: early life decides. Int J Mol Sci 2017;18:1711.
- 12. Targum SD, Nemeroff CB. The effect of early life stress on adult psychiatric disorders. Innov Clin Neurosci 2019;16:35–7
- 13. Duman RS, Sanacora G, Krystal JH. Altered connectivity in depression: GABA and glutamate neurotransmitter deficits and reversal by novel treatments. Neuron 2019;102:75–90
- 14. Isella V, Iurlaro S, Piolti R, Ferrarese C, Frattola L, Appollonio I, Melzi P, Grimaldi M. Physical anhedonia in Parkinson's disease. J Neurol Neurosurg Psychiatry 2003;74:1308–11
- 15. Strauss GP, Gold JM. A new perspective on anhedonia in schizophrenia. Am J Psychiatry 2012;169:364–73
- 16. Pizzagalli DA. Depression, stress, and anhedonia: toward a synthesis and integrated model. Annu Rev Clin Psychol 2014;10:393–423
- 17. Treadway MT, Zald DH. Reconsidering anhedonia in depression: lessons from translational neuroscience. Neurosci Biobehav Rev 2011;35:537–55
- 18. Klein DF. Endogenomorphic depression. A conceptual and terminological revision. Arch Gen Psychiatry 1974;31:447–54
- 19. Nutt DJ. Relationship of neurotransmitters to the symptoms of major depressive disorder. J Clin Psychiatry 2008;69 Suppl E1:4–7
- 20. Frederick AL, Yano H, Trifilieff P, Vishwasrao HD, Biezonski D, Mészáros J, Urizar E, Sibley DR, Kellendonk C, Sonntag KC, Graham DL, Colbran RJ, Stanwood GD, Javitch JA. Evidence against dopamine D1/D2 receptor heteromers. Mol Psychiatry 2015;20:1373–85
- 21. Floresco SB, Tse MT. Dopaminergic regulation of inhibitory and excitatory transmission in the basolateral amygdala-prefrontal cortical pathway. J Neurosci 2007;27:2045–57
- 22. Pisani A, Bonsi P, Centonze D, Calabresi P, Bernardi G. Activation of D2-like dopamine receptors reduces synaptic inputs to striatal cholinergic interneurons. J Neurosci 2000;20:Rc69
- 23. Stefani A, De Murtas M, Pisani A, Stratta F, Bonci A, Mercuri NB, Calabresi P. Electrophysiology of dopamine D-1 receptors in the basal ganglia: old facts and new perspectives. Prog Neuropsychopharmacol Biol Psychiatry 1995;19:779–93
- 24. Kebabian JW. Multiple classes of dopamine receptors in mammalian central nervous system: the involvement of dopamine-sensitive adenylyl cyclase. Life Sci 1978;23:479–83
- 25. Sidhu A. Coupling of D1 and D5 dopamine receptors to multiple G proteins: implications for understanding the diversity in receptor-G protein coupling. Mol Neurobiol 1998;16:125–34
- 26. Savasta M, Dubois A, Scatton B. Autoradiographic localization of D1 dopamine receptors in the rat brain with [3H]SCH 23390. Brain Res 1986;375:291–301
- 27. Wamsley JK, Gehlert DR, Filloux FM, Dawson TM. Comparison of the distribution of D-1 and D-2 dopamine receptors in the rat brain. J Chem Neuroanat 1989;2:119–37
- 28. Baik JH. Dopamine signaling in reward-related behaviors. Front Neural Circuits 2013;7:152
- 29. Yokoyama N, Kuno T, Furuyama S, Wang JH. Immunological approach to identify calmodulin-stimulated phosphatase isozymes from bovine brain. Mol Cell Biochem 1994;132:101–8
- 30. Stanwood GD, Artymyshyn RP, Kung MP, Kung HF, Lucki I, McGonigle P. Quantitative autoradiographic mapping of rat brain dopamine D3 binding with [(125)I]7-OH-PIPAT: evidence for the presence of D3 receptors on dopaminergic and nondopaminergic cell bodies and terminals. J Pharmacol Exp Ther 2000;295:1223–31
- 31. Liu L, van Groen T, Kadish I, Tollefsbol TO. DNA methylation impacts on learning and memory in aging. Neurobiol Aging 2009;30:549–60
- 32. Chang L, Karin M. Mammalian MAP kinase signalling cascades. Nature 2001;410:37–40
- 33. Stanwood GD. Dopamine Stress. In: Fink G (ed.) Stress: physiology, biochemistry and pathology. 3rd ed. London: Elsevier, 2019, pp.105–23
- 34. Haber SN. The place of dopamine in the cortico-basal ganglia circuit. Neuroscience 2014;282:248–57
- 35. Kapur S, Mann JJ. Role of the dopaminergic system in depression. Biol Psychiatry 1992;32:1–17
- 36. Francis TC, Lobo MK. Emerging role for nucleus accumbens medium spiny neuron subtypes in depression. Biol Psychiatry 2017;81:645–53
- 37. Nestler EJ, Carlezon WA Jr. The mesolimbic dopamine reward circuit in depression. Biol Psychiatry 2006;59:1151–9
- 38. Grace AA. Dysregulation of the dopamine system in the pathophysiology of schizophrenia and depression. Nat Rev Neurosci 2016;17:524–32
- 39. Deutch AY, Roth RH. The determinants of stress-induced activation of the prefrontal cortical dopamine system. Prog Brain Res 1990;85:367–402; discussion
- 40. Horger BA, Roth RH. The role of mesoprefrontal dopamine neurons in stress. Crit Rev Neurobiol 1996;10:395–418
- 41. Moghaddam B, Jackson M. Effect of stress on prefrontal cortex function. Neurotox Res 2004;6:73–8
- 42. Finlay JM, Zigmond MJ. The effects of stress on central dopaminergic neurons: possible clinical implications. Neurochem Res 1997;22:1387–94
- 43. Abercrombie ED, Keefe KA, DiFrischia DS, Zigmond MJ. Differential effect of stress on in vivo dopamine release in striatum, nucleus accumbens, and medial frontal cortex. J Neurochem 1989;52:1655–8
- 44. Pezze MA, Feldon J. Mesolimbic dopaminergic pathways in fear conditioning. Prog Neurobiol 2004;74:301–20
- 45. Cervo L, Samanin R. Repeated treatment with imipramine and amitriptyline reduced the immobility of rats in the swimming test by enhancing dopamine mechanisms in the nucleus accumbens. J Pharm Pharmacol 1988;40:155–6
- 46. Chen P. Optimized treatment strategy for depressive disorder. Adv Exp Med Biol 2019;1180:201–17
- 47. Keks NA, Hope J, Castle D. Lurasidone: an antipsychotic with antidepressant effects in bipolar depression? Australas Psychiatry 2016;24:289–91
- 48. Fornaro M, Fusco A, Anastasia A, Cattaneo CI, De Berardis D. Brexpiprazole for treatment-resistant major depressive disorder. Expert Opin Pharmacother 2019;20:1925–33
- 49. Mallet J, Gorwood P, Le Strat Y, Dubertret C. Major depressive disorder (MDD) and schizophrenia- addressing unmet needs with partial agonists at the D2 receptor: a review. Int J Neuropsychopharmacol 2019;22:651–64
- 50. Planchez B, Surget A, Belzung C. Animal models of major depression: drawbacks and challenges. J Neural Transm 2019;126:1383–408
- 51. Gururajan A, Reif A, Cryan JF, Slattery DA. The future of rodent models in depression research. Nat Rev Neurosci 2019;20:686–701
- 52. Wang Q, Timberlake MA 2nd, Prall K, Dwivedi Y. The recent progress in animal models of depression. Prog Neuropsychopharmacol Biol Psychiatry 2017;77:99–109
- 53. Hershenberg R, Satterthwaite TD, Daldal A, Katchmar N, Moore TM, Kable JW, Wolf DH. Diminished effort on a progressive ratio task in both unipolar and bipolar depression. J Affect Disord 2016;196:97–100
- 54. Der-Avakian A, D'Souza MS, Pizzagalli DA, Markou A. Assessment of reward responsiveness in the response bias probabilistic reward task in rats: implications for cross-species translational research. Transl Psychiatry 2013;3:e297
- 55. Kaiser RH, Treadway MT, Wooten DW, Kumar P, Goer F, Murray L, Beltzer M, Pechtel P, Whitton A, Cohen AL, Alpert NM, El Fakhri G, Normandin MD, Pizzagalli DA. Frontostriatal and dopamine markers of individual differences in reinforcement learning: a multi-modal investigation. Cereb Cortex 2018;28:4281–90
- 56. Davis GL, Stewart A, Stanwood GD, Gowrishankar R, Hahn MK, Blakely RD. Functional coding variation in the presynaptic dopamine transporter associated with neuropsychiatric disorders drives enhanced motivation and context-dependent impulsivity in mice. Behav Brain Res 2018;337:61–9
- 57. Liu MY, Yin CY, Zhu LJ, Zhu XH, Xu C, Luo CX, Chen H, Zhu DY, Zhou QG. Sucrose preference test for measurement of stress-induced anhedonia in mice. Nat Protoc 2018;13:1686–98
- 58. Olney JJ, Marshall SA, Thiele TE. Assessment of depression-like behavior and anhedonia after repeated cycles of binge-like ethanol drinking in male C57BL/6J mice. Pharmacol Biochem Behav 2018;168:1–7
- 59. Caspi A, Harrington H, Moffitt TE, Milne BJ, Poulton R. Socially isolated children 20 years later: risk of cardiovascular disease. Arch Pediatr Adolesc Med 2006;160:805–11
- 60. Uher R, Perroud N, Ng MY, Hauser J, Henigsberg N, Maier W, Mors O, Placentino A, Rietschel M, Souery D, Zagar T, Czerski PM, Jerman B, Larsen ER, Schulze TG, Zobel A, Cohen-Woods S, Pirlo K, Butler AW, Muglia P, Barnes MR, Lathrop M, Farmer A, Breen G, Aitchison KJ, Craig I, Lewis CM, McGuffin P. Genome-wide pharmacogenetics of antidepressant response in the GENDEP project. Am J Psychiatry 2010;167:555–64
- 61. Menard C, Pfau ML, Hodes GE, Russo SJ. Immune and neuroendocrine mechanisms of stress vulnerability and resilience. Neuropsychopharmacology 2017;42:62–80
- 62. Lu Q, Mouri A, Yang Y, Kunisawa K, Teshigawara T, Hirakawa M, Mori Y, Yamamoto Y, Libo Z, Nabeshima T, Saito K. Chronic unpredictable mild stress-induced behavioral changes are coupled with dopaminergic hyperfunction and serotonergic hypofunction in mouse models of depression. Behav Brain Res 2019;372:112053
- 63. Yang CR, Zhang ZG, Bai YY, Zhou HF, Zhou L, Ruan CS, Li F, Li CQ, Zheng HY, Shen LJ, Zhou XF. Foraging activity is reduced in a mouse model of depression. Neurotox Res 2014;25:235–47
- 64. Chandra R, Francis TC, Nam H, Riggs LM, Engeln M, Rudzinskas S, Konkalmatt P, Russo SJ, Turecki G, Iniguez SD, Lobo MK. Reduced Slc6a15 in nucleus accumbens D2-neurons underlies stress susceptibility. J Neurosci 2017;37:6527–38
- 65. Chaudhury D, Walsh JJ, Friedman AK, Juarez B, Ku SM, Koo JW, Ferguson D, Tsai HC, Pomeranz L, Christoffel DJ, Nectow AR, Ekstrand M, Domingos A, Mazei-Robison MS, Mouzon E, Lobo MK, Neve RL, Friedman JM, Russo SJ, Deisseroth K, Nestler EJ, Han MH. Rapid regulation of depression-related behaviours by control of midbrain dopamine neurons. Nature 2013;493:532–6
- 66. Heshmati M, Aleyasin H, Menard C, Christoffel DJ, Flanigan ME, Pfau ML, Hodes GE, Lepack AE, Bicks LK, Takahashi A, Chandra R, Turecki G, Lobo MK, Maze I, Golden SA, Russo SJ. Cell-type-specific role for nucleus accumbens neuroligin-2 in depression and stress susceptibility. Proc Natl Acad Sci U S A 2018;115:1111-6
- 67. Bergamini G, Mechtersheimer J, Azzinnari D, Sigrist H, Buerge M, Dallmann R, Freije R, Kouraki A, Opacka-Juffry J, Seifritz E, Ferger B, Suter T, Pryce CR. Chronic social stress induces peripheral and central immune activation, blunted mesolimbic dopamine function, and reduced reward-directed behaviour in mice. Neurobiol Stress 2018;8:42–56
- 68. Shimamoto A. Social defeat stress, sex, and addiction-Like behaviors. Int Rev Neurobiol 2018;140:271–313
- 69. Burstein O, Doron R. The unpredictable chronic mild stress protocol for inducing anhedonia in mice. J Vis Exp 2018;140:58184.
- 70. Hendrie CA, Pickles AR. Depression as an evolutionary adaptation: implications for the development of preclinical models. Med Hypotheses 2009;72:342–7
- 71. Tye KM, Mirzabekov JJ, Warden MR, Ferenczi EA, Tsai HC, Finkelstein J, Kim SY, Adhikari A, Thompson KR, Andalman AS, Gunaydin LA, Witten IB, Deisseroth K. Dopamine neurons modulate neural encoding and expression of depression-related behaviour. Nature 2013;493:537–41
- 72. Freund N, Thompson BS, Sonntag K, Meda S, Andersen SL. When the party is over: depressive-like states in rats following termination of cortical D1 receptor overexpression. Psychopharmacology 2016;233:1191–201
- 73. Bagot RC, Parise EM, Peña CJ, Zhang HX, Maze I, Chaudhury D, Persaud B, Cachope R, Bolaños-Guzmán CA, Cheer JF, Deisseroth K, Han MH, Nestler EJ. Ventral hippocampal afferents to the nucleus accumbens regulate susceptibility to depression. Nat Commun 2015;6:7062
- 74. de Jong JW, Afjei SA, Pollak Dorocic I, Peck JR, Liu C, Kim CK, Tian L, Deisseroth K, Lammel S. A neural circuit mechanism for encoding aversive stimuli in the mesolimbic dopamine system. Neuron 2019;101:133–51 e7
- 75. Warden MR, Selimbeyoglu A, Mirzabekov JJ, Lo M, Thompson KR, Kim SY, Adhikari A, Tye KM, Frank LM, Deisseroth K. A prefrontal cortex-brainstem neuronal projection that controls response to behavioural challenge. Nature 2012;492:428–32
- 76. Sidor MM, Spencer SM, Dzirasa K, Parekh PK, Tye KM, Warden MR, Arey RN, Enwright JF 3rd, Jacobsen JP, Kumar S, Remillard EM, Caron MG, Deisseroth K, McClung CA. Daytime spikes in dopaminergic activity drive rapid mood-cycling in mice. Mol Psychiatry 2015;20:1406–19
- 77. Miller SM, Marcotulli D, Shen A, Zweifel LS. Divergent medial amygdala projections regulate approach-avoidance conflict behavior. Nat Neurosci 2019;22:565–75
- 78. Matthews GA, Nieh EH, Vander Weele CM, Halbert SA, Pradhan RV, Yosafat AS, Glober GF, Izadmehr EM, Thomas RE, Lacy GD, Wildes CP, Ungless MA, Tye KM. Dorsal raphe dopamine neurons represent the experience of social isolation. Cell 2016;164:617–31
- 79. DeGroot SR, Zhao-Shea R, Chung L, Klenowski PM, Sun F, Molas S, Gardner PD, Li Y, Tapper AR. Midbrain dopamine controls anxietylike behavior by engaging unique interpeduncular nucleus microcircuitry. Biol Psychiatry 2020;88:855–66
- 80. Shinohara R, Taniguchi M, Ehrlich AT, Yokogawa K, Deguchi Y, Cherasse Y, Lazarus M, Urade Y, Ogawa A, Kitaoka S, Sawa A, Narumiya S, Furuyashiki T. Dopamine D1 receptor subtype mediates acute stress-induced dendritic growth in excitatory neurons of the medial prefrontal cortex and contributes to suppression of stress susceptibility in mice. Mol Psychiatry 2018;23:1717–30
- 81. Mondoloni S, Durand-de Cuttoli R, Mourot A. Cell-Specific neuropharmacology. Trends Pharmacol Sci 2019;40:696–710
- 82. Pizzagalli DA, Berretta S, Wooten D, Goer F, Pilobello KT, Kumar P, Murray L, Beltzer M, Boyer-Boiteau A, Alpert N, El Fakhri G, Mechawar N, Vitaliano G, Turecki G, Normandin M. Assessment of striatal dopamine transporter binding in individuals with major depressive disorder: in vivo positron emission tomography and postmortem evidence. JAMA Psychiatry 2019;76:854–61
- 83. Pizzagalli DA, Holmes AJ, Dillon DG, Goetz EL, Birk JL, Bogdan R, Dougherty DD, Iosifescu DV, Rauch SL, Fava M. Reduced caudate and nucleus accumbens response to rewards in unmedicated individuals with major depressive disorder. Am J Psychiatry 2009;166:702-10
- 84. Forbes EE, Hariri AR, Martin SL, Silk JS, Moyles DL, Fisher PM, Brown SM, Ryan ND, Birmaher B, Axelson DA, Dahl RE. Altered striatal activation predicting real-world positive affect in adolescent major depressive disorder. Am J Psychiatry 2009;166:64–73
- 85. Kunii Y, Zhang W, Xu Q, Hyde TM, McFadden W, Shin JH, Deep-Soboslay A, Ye T, Li C, Kleinman JE, Wang KH, Lipska BK.

CHRNA7 and CHRFAM7A mRNAs: co-localized and their expression levels altered in the postmortem dorsolateral prefrontal cortex in major psychiatric disorders. Am J Psychiatry 2015;172:1122–30

- 86. Cathomas F, Murrough JW, Nestler EJ, Han MH, Russo SJ. Neurobiology of resilience: interface between mind and body. Biol Psychiatry 2019;86:410–20
- 87. Ansorge MS, Hen R, Gingrich JA. Neurodevelopmental origins of depressive disorders. Curr Opin Pharmacol 2007;7:8–17
- 88. Suri D, Teixeira CM, Cagliostro MK, Mahadevia D, Ansorge MS. Monoamine-sensitive developmental periods impacting adult emotional and cognitive behaviors. Neuropsychopharmacology 2015;40:88–112
- 89. Courtin J, Chaudun F, Rozeske RR, Karalis N, Gonzalez-Campo C, Wurtz H, Abdi A, Baufreton J, Bienvenu TC, Herry C. Prefrontal parvalbumin interneurons shape neuronal activity to drive fear expression. Nature 2014;505:92–6
- 90. Graham DL, Durai HH, Garden JD, Cohen EL, Echevarria FD, Stanwood GD. Loss of dopamine D2 receptors increases parvalbumin-positive interneurons in the anterior cingulate cortex. ACS Chem Neurosci 2015;6:297–305
- 91. Northoff G, Sibille E. Why are cortical GABA neurons relevant to internal focus in depression? A cross-level model linking cellular, biochemical and neural network findings. Mol Psychiatry 2014;19:966–77
- 92. Sauer JF, Struber M, Bartos M. Impaired fast-spiking interneuron function in a genetic mouse model of depression. eLife 2015;4:e04979.
- 93. Sibille E, Morris HM, Kota RS, Lewis DA. GABA-related transcripts in the dorsolateral prefrontal cortex in mood disorders. Int J Neuropsychopharmacol 2011;14:721–34
- 94. Flames N, Marin O. Developmental mechanisms underlying the generation of cortical interneuron diversity. Neuron 2005;46:377–81
- 95. Jones EG. The origins of cortical interneurons: mouse versus monkey and human. Cereb Cortex 2009;19:1953–6
- 96. Wamsley B, Fishell G. Genetic and activity-dependent mechanisms underlying interneuron diversity. Nat Rev Neurosci 2017;18:299–309
- 97. Xu Q, Cobos I, De La Cruz E, Rubenstein JL, Anderson SA. Origins of cortical interneuron subtypes. J Neurosci 2004;24:2612–22
- 98. Gaspar P, Bloch B, Le Moine C. D1 and D2 receptor gene expression in the rat frontal cortex: cellular localization in different classes of efferent neurons. Eur J Neurosci 1995;7:1050–63
- 99. Muly EC 3rd, Szigeti K, Goldman-Rakic PS. D1 receptor in interneurons of macaque prefrontal cortex: distribution and subcellular localization. J Neurosci 1998;18:10553–65
- 100. Thompson BL, Stanwood GD, Levitt P. Specificity of prenatal cocaine exposure effects on cortical interneurons is independent from dopamine D1 receptor co-localization. J Chem Neuroanat 2010;39:228–34
- 101. Xu L, Zhang XH. Distribution of D1 and D2-dopamine receptors in calcium-binding-protein expressing interneurons in rat anterior cingulate cortex. Sheng Li Xue Bao 2015;67:163–72.
- 102. Money KM, Stanwood GD. Developmental origins of brain disorders: roles for dopamine. Front Cell Neurosci 2013;7:260
- 103. Frederick AL, Stanwood GD. Drugs, biogenic amine targets and the developing brain. Dev Neurosci 2009;31:7–22
- 104. Thompson BL, Levitt P, Stanwood GD. Prenatal exposure to drugs: effects on brain development and implications for policy and education. Nat Rev Neurosci 2009;10:303–12
- 105. Francis TC, Gaynor A, Chandra R, Fox ME, Lobo MK. The selective RhoA inhibitor rhosin promotes stress resiliency through enhancing D1-medium spiny neuron plasticity and reducing hyperexcitability. Biol Psychiatry 2019;85:1001–10
- 106. Jones LB, Stanwood GD, Reinoso BS, Washington RA, Wang HY, Friedman E, Levitt P. In utero cocaine-induced dysfunction of dopamine D1 receptor signaling and abnormal differentiation of cerebral cortical neurons. J Neurosci 2000;20:4606–14
- 107. Song ZM, Undie AS, Koh PO, Fang YY, Zhang L, Dracheva S, Sealfon SC, Lidow MS. D1 dopamine receptor regulation of microtubuleassociated protein-2 phosphorylation in developing cerebral cortical neurons. J Neurosci 2002;22:6092–105
- 108. Stanwood GD, Parlaman JP, Levitt P. Anatomical abnormalities in dopaminoceptive regions of the cerebral cortex of dopamine D1 receptor mutant mice. J Comp Neurol 2005;487:270–82
- 109. Stanwood GD, Parlaman JP, Levitt P. Genetic or pharmacological inactivation of the dopamine D1 receptor differentially alters the expression of regulator of G-protein signalling (Rgs) transcripts. Eur J Neurosci 2006;24:806–18
- 110. Stanwood GD, Washington RA, Levitt P. Identification of a sensitive period of prenatal cocaine exposure that alters the development of the anterior cingulate cortex. Cereb Cortex 2001;11:430–40
- 111. Crandall JE, McCarthy DM, Araki KY, Sims JR, Ren JQ, Bhide PG. Dopamine receptor activation modulates GABA neuron migration from the basal forebrain to the cerebral cortex. J Neurosci 2007;27:3813–22
- 112. Ohtani N, Goto T, Waeber C, Bhide PG. Dopamine modulates cell cycle in the lateral ganglionic eminence. J Neurosci 2003;23:2840–50
- 113. Brown RW, Flanigan TJ, Thompson KN, Thacker SK, Schaefer TL, Williams MT. Neonatal quinpirole treatment impairs Morris water task performance in early postweanling rats: relationship to increases in corticosterone and decreases in neurotrophic factors. Biol Psychiatry 2004;56:161–8
- 114. Pruessner JC, Champagne F, Meaney MJ, Dagher A. Dopamine release in response to a psychological stress in humans and its relationship to

early life maternal care: a positron emission tomography study using [11C]raclopride. J Neurosci 2004;24:2825–31

- 115. Pruessner JC, Dedovic K, Pruessner M, Lord C, Buss C, Collins L, Dagher A, Lupien SJ. Stress regulation in the central nervous system: evidence from structural and functional neuroimaging studies in human populations – 2008 Curt Richter Award Winner. Psychoneuroendocrinology 2010;35:179–91
- 116. Teicher MH, Andersen SL, Polcari A, Anderson CM, Navalta CP, Kim DM. The neurobiological consequences of early stress and childhood maltreatment. Neurosci Biobehav Rev 2003;27:33–44
- 117. Burke AR, Miczek KA. Stress in adolescence and drugs of abuse in rodent models: role of dopamine, CRF, and HPA axis. Psychopharmacology 2014;231:1557–80
- 118. Ernst M, Romeo RD, Andersen SL. Neurobiology of the development of motivated behaviors in adolescence: a window into a neural systems model. Pharmacol Biochem Behav 2009;93:199–211
- 119. Paus T, Keshavan M, Giedd JN. Why do many psychiatric disorders emerge during adolescence? Nat Rev Neurosci 2008;9:947–57
- 120. Mastwal S, Ye Y, Ren M, Jimenez DV, Martinowich K, Gerfen CR, Wang KH. Phasic dopamine neuron activity elicits unique mesofrontal plasticity in adolescence. J Neurosci 2014;34:9484–96