

Role of hyperoxic treatment in cancer

Sei W Kim¹ , In K Kim^{1,2} and Sang H Lee^{1,2}

¹Division of Pulmonary, Critical Care and Sleep Medicine, Department of Internal Medicine, Eunpyeong St. Mary's Hospital, College of Medicine, The Catholic University of Korea, Seoul 03312, Republic of Korea; ²Cancer Research Institute, College of Medicine, The Catholic University of Korea, Seoul 06591, Republic of Korea

Corresponding author: Sang H Lee. Email: mdlee@catholic.ac.kr

Impact statement

Tumor hypoxia promotes cancer cell aggressiveness, and is strongly associated with poor prognosis across multiple tumor types. The hypoxic microenvironments inside tumors also limit the effectiveness of radiotherapy, chemotherapy, and immunotherapy. Several approaches to eliminate hypoxic state in tumors have been proposed to delay cancer progression and improve therapeutic efficacies. This review will summarize current knowledge on hyperoxia, used alone or in combination with other therapeutic modalities, in cancer treatment. Molecular mechanisms and undesired side effects of hyperoxia will also be discussed.

Abstract

The occurrence of hypoxia is common in many solid tumors, and it enhances aggressive features of cancer such as cell survival, angiogenesis, and metastasis while minimizing the efficacies of chemotherapy and radiotherapy. Hypoxia also plays a pivotal role in regulating immune cell function which is important for immunotherapy. Hypoxia-inducible factor has been suggested as a master regulator of tumor cell adaptation to the hypoxic microenvironment. Currently, several approaches have been proposed to eliminate the hypoxic state in tumors for delaying cancer progression and improving therapeutic efficacy. In this review, we summarize current findings on the relevance of hyperoxia-based therapeutics for cancer treatment. Accumulating evidence indicates that hyperoxic therapy inhibits tumor growth and increases treatment efficacy. Primary antitumor effect of hyperoxic therapy may be due to the reversal of tumor hypoxia and the generation of reactive oxygen species. Restoring immune function is also suggested as a potential mechanism. Hyperoxic therapy can also

cause cellular injury and organ dysfunction. In conclusion, overcoming tumor hypoxia is a major problem that needs to be solved. Further studies to standardize and personalize hyperoxia therapy according to the type of cancer, stage, and comorbidities are needed.

Keywords: Neoplasms, hyperoxia, oxygen, reactive oxygen species, oxidative stress, antitumor effect

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Introduction

Oxygen is an indispensable element for cells in our body to fulfill energy requirement from aerobic metabolism. Therefore, reduced oxygen levels, hypoxia, may produce cell death and resulting organ dysfunction as occurs in heart attacks or strokes. Hypoxia commonly arises in the core of most solid tumors as a result of an inadequate supply of oxygen from abnormal vasculature, and an increased oxygen demand from changes in tumor metabolism.¹ Severity of tumor hypoxia varies depending on tissue of origin and tumor size, and median oxygen levels in untreated tumors are frequently less than 2% in comparison to the normal human tissues (5%).^{2,3} The hypoxic regions within the tumor are heterogeneously distributed and may even be located near to vessels.⁴

Traditionally, hypoxia was thought of as a factor limiting the cancer growth by reducing the ability of cells to divide.⁵ However, increasing evidence indicates that tumor hypoxia plays an important role in cancer progression. When tumor cells adapt to the imbalance between oxygen supply and demand, malignant features of solid tumors such as resistance to cell death, angiogenesis, and metastasis were invariably enhanced.^{6,7} Emerging features of cancer development such as genome instability, the enrichment of cancer stem cells, and aberrant exosomal secretion were also suggested as hallmarks of tumor hypoxia.^{8–10} Overall, hypoxic microenvironment in tumor promotes acquisition of the more aggressive cancer phenotypes and thus is associated with poor prognosis.^{11,12} In addition, tumor hypoxia has been considered as one of the biggest barriers to treating cancer because it renders tumors more

resistant to surgery, radiotherapy, chemotherapy, and immunotherapy.^{13–16} In this regard, it is widely accepted that hypoxia is a potential cancer-specific target,^{11,17} and several approaches targeting tumor hypoxia for cancer therapy such as erythropoietin, nicotinamide, hypoxia-activated prodrugs, and nanocarriers have been proposed.^{14,18}

Numerous studies also suggest increasing oxygen content using hyperoxia as a promising therapeutic modality to reverse tumor hypoxia as well as resistance to cancer therapeutics.^{5,19–21} Recently, we also identified anti-cancer effect of hyperoxia and its mechanism of action in mouse lung cancer.^{22,23} However, despite of growing interest and clinical trials, the application of hyperoxia to cancer treatment is still in its nascent stage. Here, we will focus on the recent insights into the understanding of hyperoxia-mediated cancer treatment.

Hyperoxic therapy in cancer

Oxygen therapy is a type of treatment to enhance the amount of dissolved oxygen in the plasma by breathing supplemental oxygen, thereby increasing O₂ delivery to body tissue. For hyperoxygenation, 100% oxygen is administered to a patient either at ambient atmospheric pressure (normobaric oxygen, NBO) or under hyperbaric conditions usually in a pressurized chamber (hyperbaric oxygen, HBO). Oxygen therapy has long been regarded as integral to the management of various medical conditions for centuries. For cancer treatment, there is still no evidence that it is effective in treating cancer.²⁴ However, despite ongoing controversy, interest in applying hyperoxic therapy as adjuvant for cancer treatment is rapidly growing. For example, in Pubmed search concerning oxygen therapy and cancer, since the first article in 1951,²⁵ more than 20,000 were found. A brief summary of hyperoxic therapies on cancer is listed in Table 1.

In addition to oxygen breathing, although relatively uncommon, other forms of oxygen therapy such as oxygen-containing substances (i.e. ozone or hydrogen peroxide), and prodrugs that are activated only in the hypoxic tumor environment also have been tried for cancer treatment.^{32–34}

Hyperbaric oxygen therapy

HBO treatment is breathing 100% oxygen at higher atmospheric pressure usually between 1.5 and 3 atm which enable the lungs to receive more oxygen up to three times than under normal air pressure, resulting in immediate saturation of plasma with oxygen. In normal subject, compared to normobaric air, HBO at 3 atm increases both arterial (from 100 to 200 mmHg) and tissue (from 55 to 500 mmHg) oxygen tensions (PO₂).³⁵ Similarly, the PO₂ level was significantly increased in tumor tissue by HBO exposure, demonstrating three to four times greater hyperoxygenation effect in tumor tissue than NBO.^{36,37} Therefore, HBO rather than NBO has been preferentially used to eliminate poorly oxygenated regions of tumor which play a major role in tumor development and

resistance to other therapeutic modalities. In addition, HBO therapy is considered safe and well tolerated, and side effects are rare.³⁸ Although several studies showed the favorable outcomes of HBO by itself in cancer treatment,^{5,28,39} HBO alone gives a limited curative effects, and it even enhances tumor growth.^{19,40} Accordingly, HBO is preferentially used as an adjuvant treatment for enhancing tumor sensitivity and decreasing complications of other therapies.³⁴

Normobaric oxygen therapy

NBO therapy is a routine adjuvant oxygenation intervention supplied by nasal cannula or facemask under ambient pressure. For cancer treatment, NBO is an attractive alternative to HBO due to its ease of administration and lower complication in actual clinical practice.⁴¹ However, when compared to HBO, effect of NBO on arterial and tissue oxygenation is much weaker. Arterial PO₂ in aorta has increased only 4-fold after exposure to NBO (345 vs. 84.1 mmHg).⁴² In animal study, most of tumors were nearly anoxic (PO₂ <1 mmHg), but NBO immediately increased PO₂ (mean >25 mmHg) and remained elevated during gas exposure.⁴³ Accordingly, NBO treatment significantly retarded tumor growth.^{39,44} We also previously showed the NBO inhibits lung cancer in *in vivo* and *in vitro* through reactive oxygen species (ROS) generation and apoptosis.²³

Molecular mechanisms: Hypoxia-inducible factor 1 α

Hypoxia

When oxygen levels in tumor microenvironment are dropping, transcriptional induction of a series of genes necessary for maintaining cell survival/proliferation and promoting more aggressive features occurs in tumor cells. These oxygen-dependent responses are tightly regulated by HIF-1 α , the master transcriptional regulator of the hypoxic response as well as a representative endogenous biomarker for hypoxia.⁴⁵ HIF-1 α is induced by hypoxia through post-translational modification, and it binds to specific recognition sequences in the genome to increase the expression of HIF-1 α target genes.⁴⁶ Since the seminal discovery of HIF in the early 1990s by Gregg Semenza, a Nobel Laureate in Physiology or Medicine for 2019,⁴⁷ thousands of genes are identified as direct targets of HIF-1 α ,⁴⁸ and therefore a myriad of changes associated with tumor aggravation occur in hypoxic tumor cells including angiogenesis and oxygen supply, stemness/self-renewal, proliferation, epithelial to mesenchymal transition, metastasis and invasion, redox homeostasis, anti-apoptosis, and metabolic reprogramming.^{49,50} Metabolic reprogramming from oxidative phosphorylation to accelerated glycolysis in cancer cells is also known to be mediated via HIF-1 α .⁵¹ In addition, HIF-1 α contributes to the development of tumor resistance to therapeutic approaches and serves as a promising biomarker.^{52,53} For example, HIF-1 α limits T cell recognition of tumor cells by downregulating MHC class I

Table 1. Summary of studies on cancer with hyperoxic therapies.

References	Year	Type of tumor	Type of hyperoxic treatment	Combination treatment	Summary of the study
Moen et al. ²⁶	2009	DMBA-induced mammary tumor mouse model	2 bar, pO ₂ = 2 bar, 4 times (day 1, 4, 7 and 10) exposures of 90 min	5-FU	HBO increases the uptake of [³ H]-5FU
Kawasoe et al. ²⁷	2009	Osteosarcoma LM8 cells mouse model	2.5 bar, pO ₂ = 2.5 bar, 60 min, 5 times a week until 5 weeks	Carboplatin	HBO or HBO with carboplatin inhibits tumor growth and lung metastasis, synergistically
Selvendiran et al. ²⁸	2010	A2780 ovarian xenograft tumor mouse model	100% O ₂ 2 atm for 90 min, 21 days	Cisplatin	Reduction of tumor volume. No significance between HBO and cisplatin-treated group
Sun et al. ²⁹	2012	Human glioblastoma multiforme cells (D54, U87)	40%, 80% O ₂ , 72 h	Temozolomide	NBO enhanced TMZ toxicity in GBM cells
Lee et al. ³⁰	2014	Human glioblastoma multiforme cells (D54, U87, U251)	40% O ₂ , 24, 48 and 72 h	Temozolomide	NBO enhanced the sensitivity to temozolomide in chemosensitive and -resistant GBM cells
Hatfield et al. ³¹	2016	MCA205 fibrosarcoma mouse model	60% O ₂ , 72 h and 11 days	–	NBO reverses the hypoxia-A2-adenosinergic immunosuppression during acute inflammation
Ytterian Sletta et al. ²¹	2017	Breast cancer mouse model	2.5 bar, pO ₂ = 2.5 bar, 90 min, every third day until day 16	5-FU	Suppressed tumor growth and metastatic lesions, HBO does not enhance the 5-FU efficacy
Kim et al. ²³	2018	Lewis lung carcinoma cell injected mouse model	24 h NBO (95% O ₂)/normoxia cycle for two weeks	–	NBO inhibits the progression of lung cancer by inducing apoptosis
Lee et al. ²²	2018	benzo[<i>a</i>]pyrene -induced lung tumorigenesis mouse model	95% O ₂ for 3 h/day, days 21–28	Carboplatin	Intermittent NBO with carboplatin displays a synergistic tumoricidal effect
Qian et al. ²⁰	2019	triple-negative breast cancer mouse model	60% O ₂ , day 7–28	–	NBO reverses immunosuppression and control the extend of lung metastases

molecule expression.⁵⁴ It also induces multidrug resistance, the major cause of chemotherapy failure, by inducing multidrug resistance-associated protein 1 in cancer cells.⁵⁵ Accordingly, in cancer patients, protein levels of HIF-1 α in solid tumors are considered as a critical prognostic factor.⁵⁶ Based on these findings, HIF-1 α has become targets for developing novel cancer therapeutics. However, no agents directly inhibiting HIF-1 α have been approved for treating cancer patients.⁵⁷

Hyperoxia

Many reports suggest various beneficial effects of hyperoxia on hypoxic tumor. Hyperoxic breathing of 60% O₂ recovers oxygen homeostasis in tumor microenvironment hypoxia to normoxia, inhibits survival/proliferation, stemness, and immune escape of cancer cells, resensitizes chemoresistance, leading to tumor regression.^{29,58,59} We also reported anti-cancer effects of hyperoxia, alone or in combination with a chemotherapeutic drug carboplatin, on mouse lung cancer; normobaric hyperoxia significantly induces oxidative stress and apoptosis in tumor tissue and reduces tumor mass and migration/invasion.^{22,23} Differential response of cancer and normal cells to hyperoxia has also provided a treatment rationale of hyperoxia in cancer treatment. Basal antioxidant defense levels are aberrant in tumor cells, thus an increased oxidative stress has been observed. Since the superoxide dismutase activity in most tumors are lower than normal tissue, tumor cells show higher susceptibility to increased ROS activity induced by hyperoxia.⁶⁰ Synthesis of glutathione (GSH), an antioxidant preventing ROS-induced damage, increases proportional to ambient oxygen tension, but not in cancer cells.⁶¹ In consistent with these findings, we demonstrated that NBO showed anti-tumor activity in lung tumor cells but not in normal lung cells.²³ In addition, hyperoxia is angiogenic in normal tissues, but anti-angiogenic in tumor tissues.⁶²

Anti-cancer mechanisms of hyperoxia can also be inferred indirectly from clinical and experimental findings. Roles of HIF-1 α in hypoxia-mediated cancer development are widely acknowledged, and hyperoxic treatment has long been used to cure clinical disorders such as hypoxia or ischemic diseases by increasing oxygen delivery to oxygen-deficient tissues. We can also take a hint from well-known mechanisms for the induction and regulation of HIF-1 α : HIF-1 α protein is subject to degradation through an oxygen-dependent ubiquitination,⁶³ and hypoxia-induced HIF-1 α protein is rapidly decayed within 5 min upon exposure to normoxia (20% O₂).⁶⁴ In addition, transcriptional activity of HIF-1 α is enhanced by hypoxia-induced ROS,⁶⁵ and inhibited by oxygen in nonhypoxic cells.⁶⁶ Taken these findings together, it seems reasonable that therapeutic effects of hyperoxia on hypoxic tumor are mediated through not direct regulation of HIF-1 α but primarily the reversal of hypoxia and the attenuation of the HIF-mediated effects.

However, despite of accumulating evidence supporting this idea, many reports also indicate HIF-1 α -independent

effects of hyperoxia on tumor. For example, both normoxia and hyperoxia induces higher levels of HIF-1 α than hypoxia in tumors, but tumor grows faster in hypoxia group, suggesting that signaling pathways other than HIF-1 α driven response may play important roles for *in vivo* cancer cell proliferation.⁶⁷ Recent report also showed that HIF-1 α levels in tumor cells was significantly downregulated in hyperoxia (60% O₂) than normoxia (20% O₂), indicating that alternative mechanisms other than simple reversal of tumor hypoxia to normoxia underlie anti-tumor effects of hyperoxia.⁵⁹ Coincidentally, Rocco *et al.*⁶⁸ suggest that relative changes of oxygen availability rather than steady state hypoxic or hyperoxic conditions play an important role in HIF transcriptional effects. In addition, hyperoxia activates HIF-1 α overexpression through the activation of Src oncogene, and also inhibits the stability of HIF-1 α by reducing ROS formation.^{65,69,70} Despite numerous attempts, mechanisms underlying hyperoxia-mediated anti-cancer activity remain to be elucidated.

Side effects

Hyperoxic therapy is usually well tolerated with an acceptable rate of complications; however, as with all medical treatments, it also includes medical risks. Dependent on the type of hyperoxic therapy, patients may experience two prominent side effects due to exposure to high levels of oxygen (oxygen toxicity) or high atmospheric pressure (barotrauma). As shown above, PO₂ used in HBO therapy is much higher than that of NBO, thus oxygen toxicity is more common in patients exposed to HBO. In addition, only HBO therapy is carried out in a hyperbaric chamber, thus barotrauma occurs only in patients taking HBO therapy.

Oxygen toxicity

Exposure to high concentration of oxygen is well known cause of cell damage. For example, oxygen, at concentrations of 95% or more, is severely cytotoxic to the pulmonary cells of many animal species, including humans.⁷¹ Humans appear more resistant to oxygen-induced damage and the risk of hyperoxic acute lung injury is minimal when the FiO₂ is ≤ 0.6 ⁷² but continuous exposure to elevated levels of oxygen may cause oxygen toxicity. Major organs subject to oxygen toxicity are lungs (hyperoxic acute lung injury), central nervous system (loss of consciousness and oxygen toxicity seizure), and eye (hyperoxic myopia and cataract).⁷³⁻⁷⁵ Oxygen toxicity is believed to be mediated primarily by a production of ROS at levels exceeding the capacity of antioxidant defence mechanisms.⁷⁶ Following HBO treatment, ROS increased to about 2.14–2.44 fold in mitochondria and 1.32–1.42 fold in whole cell.⁷⁷ Reduction of antioxidant enzymes such as superoxide dismutase and glutathione by HBO, but not by NBO, also contributes to increased oxygen toxicity.⁷⁸ Excessive ROS can damage all essential macromolecules, including nucleic acids, lipids and proteins, leading to an overall progressive decline in physiological function.⁷⁹ For example, protein oxidation

and nitrosylation can impair a wide variety of enzymatic processes and growth factors that can result in marked cellular dysfunction.⁸⁰ Lipid peroxidation activates apoptosis through activation of sphingomyelinase and release of ceramide.⁸¹ Nucleic acid oxidation has been linked with aging and DNA strand breaks, leading to necrosis and/or apoptosis.⁸² Therefore, HBO-driven ROS can display many harmful activities including the induction of DNA damage, cell death, cellular senescence, and deleterious inflammatory response which in turn exacerbates oxidative toxicity and tissue damage.^{83,84} Several mechanisms involved in hyperoxia-induced oxygen toxicity have been proposed. For example, ROS stimulates signaling pathways mediated via protein kinases (Akt, MAPK and PKC), resulting in activation of transcription factors (Nrf2, NF- κ B, and AP-1) responsible for cell death and inflammation.⁸⁵ In addition, ROS-independent mechanisms such as the induction of apoptosis by direct activation of Bax, Bak, or FAS,⁸⁶ chemokine receptor CXCR2-mediated tissue inflammation,⁸⁷ and toll-like receptor-linked cell damage⁸⁸ also have been suggested.

Barotrauma and other complications

In HBO therapy, unlike to NBO, patients are exposed to the high atmospheric pressure in hyperbaric chamber. Barotrauma, pressure-induced injury, is caused by inability to equalize pressure between the environment and the air-filled space in the body such as lungs, ear, sinuses, eyes, and teeth are concurrently at risk. The most common type of barotrauma (>17%) is middle ear barotrauma which can lead to permanent hearing loss and vertigo.⁸⁹

Some patients can develop a feeling of claustrophobia, the fear of being enclosed in small spaces with no escape, due to the confined nature of hyperbaric chamber. HBO therapy also causes mild increase in blood pressure in both hypertensive and non-hypertensive patients, and hypoglycemia can occur in diabetic patients.⁹⁰ In addition, NBO increases pulmonary metastasis of tumor⁹¹ and inhibits glucose-induced insulin release.⁹²

Because hyperoxia, unlike hypoxia, is a man-made condition, specific adaptive response to hyperoxia has not been evolved in humans. To enhance safety and to prevent side-effects of HBO therapy in cancer treatment as well as other clinical trials, further investigation to maximize therapeutic efficacy and minimize complications by standardizing therapy protocol in particular with regard to pressure and duration is required.

Role of hyperoxia treatment in chemotherapy

Hypoxia reduces the sensitivity of cancer to chemotherapy.⁶ In addition, hypoxic cells do not receive sufficient chemotherapeutic agents due to distance from the capillary and because of abnormal vascularization of tumors. Hypoxia remotes resistance through the HIF-1-mediated upregulation of different genes and signaling pathways.⁹³ Hypoxia-induced drug resistance is also explained by inhibition of apoptotic pathways^{23,94} and increased intracellular drug efflux.⁹⁵ Currently, respiratory hyperoxia is mainly

used for the treatment of hypoxic tissue damage. Also, hyperoxia has also been shown to improve the treatment efficacy of chemotherapy in animal models.^{20,22,26} From our previous study, NBO therapy was found to be tumoricidal and NBO with carboplatin exhibited a synergistic antitumor effect on B[a]P-induced lung cancers in mice.^{22,23} Oxidative stress and its effects on DNA are increased following exposure to hyperoxia and even more with chemotherapy, and this may lead to apoptosis of lung tumors. Lee *et al.*³⁰ showed that NBO treatment resensitizes chemoresistant glioblastoma cells to temozolomide through unfolded protein response.

Other studies were mostly performed under HBO treatment. Moen *et al.*²⁶ showed that HBO treatment increases the uptake of 5-fluorouracil in mammary tumors for the duration of, and immediately after, HBO treatment. Kawasoe *et al.*²⁷ showed significant suppression of osteosarcoma with HBO plus carboplatin compared with monotherapy, both in *in vitro* and *in vivo*. Other studies also showed increased efficacy against variety of malignancies with combination of HBO and chemotherapy.^{28,96,97} However, precise mechanisms are not known until now and more standard combination treatment protocols are needed. In addition, a combination of HBO treatment and particular chemotherapeutic agents (doxorubicin, bleomycin, and disulfiram) may cause potential toxicity⁹⁸ because it can potentiate oxygen-related serious organ damage.^{99–101} However, studies showing conflicting results also exist.^{102,103}

Role of hyperoxic treatment in radiation therapy

The primary mechanism of radiation therapy is creation of ROS, which in turn induces cell death by the mechanisms including apoptosis, necrosis, autophagy, and senescence.¹⁰⁴ In hypoxic state, DNA radicals are repaired by abstracting hydrogen from sulfhydryl group present in protein.¹ Since oxygen is required for ROS generation, hypoxic tumors are resistant to the cytotoxic effects of radiotherapy.¹⁰⁵ HIF-1 plays a role in radioresistance of a tumor by up-regulating downstream genes, which are involved in apoptosis, metabolism, proliferation, and neovascularization.¹⁰⁶ In general, cells irradiated under normal oxygenated conditions are two- to three-fold more radiosensitive than cells irradiated under hypoxic or anoxic conditions.^{107,108} Several human studies reported significant improvement of survival and local tumor control in patients with cancer treated with radiotherapy and HBO.^{109–111} However, other studies suggest a high rate of complications from combination of HBO and radiotherapy, including severe tissue radiation injury and seizures.^{34,110} Concerning this matter, beneficial effects of NBO on tumor radiosensitivity have been also reported.^{112,113} In addition, both the extent and the timing of this hyperoxic therapy are variable. From systematic reviews in patients with high grade gliomas, radiation therapy after HBO treatment was tolerated and beneficial.¹¹¹

Role of hyperoxic treatment in immunotherapy

Oxygen tension directly affects immune cell function, and thus hypoxia can cause immunosuppression and/or immune dysfunction.¹¹⁴ It is widely appreciated that hypoxic tumor microenvironment negatively affects anti-tumor immune responses, and also is responsible for resistance to immunotherapy.^{16,115} Many mechanisms underlying hypoxia-induced immunomodulation in cancer have been suggested.¹¹⁶ Extracellular adenosine, a potent immunosuppressive metabolite, is increased in hypoxic conditions, and controlled by two cell surface nucleotidases; CD39 and CD73 in tumors,^{117,118} providing evidence that the adenosine-dependent immunoregulation is important for hypoxia-mediated immunosuppression. TGF- β , a potent immunosuppressive cytokine, is upregulated in tumor cells after culturing in hypoxic conditions.¹¹⁹ Hypoxia also increases the accumulation of intracellular adenosine by HIF-1 α -dependent mechanism, resulting in the elevation of extracellular adenosine independent of CD39/CD73.^{120,121} Programmed cell death-1 (PD-1) and programmed death-ligand 1 (PD-L1), important players in immune checkpoint pathways, are also regulated oxygen-dependently in the tumor microenvironment. Hypoxia increases PD-L1 expression which induces cancer cell resistance to T-cell dependent cytotoxicity.^{122,123} Exposure to adenosine or the activation of its receptors in T cells also downregulates T cell activities by inducing PD-1 and cytotoxic T-lymphocyte-associated protein-4 (CTLA-4) expression.^{124,125} Activities of myeloid-derived suppressor cells (MDSCs), typical immunosuppressive cells in the tumor microenvironment, are also regulated by hypoxia and extracellular adenosine.^{126,127}

Based on these findings, hyperoxia therapy has been attempted to restore hypoxia-induced impairment of immune function in cancer cells, in particular by downregulation of immune checkpoint pathways.³¹ In mice, 60% oxygen efficiently reduced tumor burden only in wild-type mice, but not in immunocompromised mice, indicating the involvement of immune response in anti-tumor activities of hyperoxia.³¹ Hyperoxia-induced alleviation of hypoxia reduces the levels of immunosuppressive molecules such as adenosine, TGF- β and PD-L1 in the tumor, and enhances anti-tumor immune responses.^{31,58,59} Population of typical immunosuppressive immune cells such as MDSCs and Treg cells in the tumor microenvironment is also decreased by hyperoxia therapy.^{20,31} Recently, Wang *et al.*⁵⁹ reported that hyperoxia reduces stemness of colorectal cancer cells through the inhibition of hypoxia-mediated production of exosome from granulocytic MDSCs.

The role of cancer immunotherapy has become increasingly important compared to traditional cancer treatments. Accumulating evidence indicates that attenuation of immunosuppressive activity in the tumor microenvironment by regulating immune checkpoints is the key factor for the success of cancer immunotherapy.¹²⁸ In this regard, it is intriguing that the inhibition of immune checkpoints pathways is the main mechanism underlying anti-tumor activity of hyperoxia.¹¹⁴ Although several immune regulators and molecular mechanisms involved in hyperoxia-

mediated antitumor activities have been identified, precise adjunctive role for hyperoxia in cancer immunotherapy still remains unclear.

Future directions and conclusion

Tumor hypoxia is a major treatment target for effective cancer therapy and inhibition of cancer progression. Hyperoxia therapy has been suggested to reverse cancer hypoxia, and it is more often used as an adjunctive treatment for cancer treatment along with other therapeutic modalities. Currently, no standard protocols for hyperoxic tumor therapy are approved. In case of combining hyperoxia with radiation, hyperoxic periods are relatively short and may not have significant side effects,³⁴ but in other cases where oxygen is administered over a long period, hyperoxia can cause cellular injury and organ dysfunction. Further, randomized, large, and well-organized clinical research could reinforce the use of hyperoxia therapy in the clinical setting for cancer treatment with minimal complications. Furthermore, more personalized approach according to the type of cancer and comorbidities are needed. Hypoxia-activated prodrugs or HIF inhibitors are suggested as possible alternatives to hyperoxic cancer therapy. The number of preclinical and clinical trials targeting low-oxygen tumor compartments via hypoxia-activated prodrugs is increasing.¹²⁹ In addition, combinations of clinical immunotherapy and immunomodulation from HIF inhibitor have a possibility of being powerful treatment option.¹³⁰ In conclusion, overcoming tumor hypoxia is an urgent problem to be solved for effective treatment of cancer patients. Further research is required to determine the mechanism for the role of oxygen tension in cancer progress and treatment, and to develop standard protocols for increasing efficacies and safety of hyperoxia therapy in cancer.

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ORCID iD

Sei W Kim  <https://orcid.org/0000-0002-2798-421X>

REFERENCES

- Brown JM, Wilson WR. Exploiting tumour hypoxia in cancer treatment. *Nat Rev Cancer* 2004;**4**:437–47
- Li XF, Carlin S, Urano M, Russell J, Ling CC, O'Donoghue JA. Visualization of hypoxia in microscopic tumors by immunofluorescent microscopy. *Cancer Res* 2007;**67**:7646–53
- McKeown SR. Defining normoxia, physoxia and hypoxia in tumours-implications for treatment response. *Br J Radiol* 2014;**87**:1–12
- Vaupel P, Mayer A. Hypoxia in cancer: significance and impact on clinical outcome. *Cancer Metastasis Rev* 2007;**26**:225–39
- Moen I, Stuhr LE. Hyperbaric oxygen therapy and cancer – a review. *Target Oncol* 2012;**7**:233–42
- Jing X, Yang F, Shao C, Wei K, Xie M, Shen H, Shu Y. Role of hypoxia in cancer therapy by regulating the tumor microenvironment. *Mol Cancer* 2019;**18**:157–72
- Rankin EB, Giaccia AJ. Hypoxic control of metastasis. *Science* 2016;**352**:175–80
- Bhandari V, Hoey C, Liu LY, Lalonde E, Ray J, Livingstone J, Lesurf R, Shiah YJ, Vujcic T, Huang X, Espiritu SMG, Heisler LE, Yousif F, Huang V, Yamaguchi TN, Yao CQ, Sabelnykova VY, Fraser M, Chua MLK, van der Kwast T, Liu SK, Boutros PC, Bristow RG. Molecular landmarks of tumor hypoxia across cancer types. *Nat Genet* 2019;**51**:308–18
- Kahlert UD, Mooney SM, Natsumeda M, Steiger HJ, Maciaczyk J. Targeting cancer stem-like cells in glioblastoma and colorectal cancer through metabolic pathways. *Int J Cancer* 2017;**140**:10–22
- Choudhry H, Harris AL. Advances in hypoxia-inducible factor biology. *Cell Metab* 2018;**27**:281–98
- Wilson WR, Hay MP. Targeting hypoxia in cancer therapy. *Nat Rev Cancer* 2011;**11**:393–410
- Campbell EJ, Dachs GU, Morrin HR, Davey VC, Robinson BA, Vissers M. Activation of the hypoxia pathway in breast cancer tissue and patient survival are inversely associated with tumor ascorbate levels. *BMC Cancer* 2019;**19**:307
- Dietl B, Marienhagen J, Schafer C, Kolbl O. The prognostic value of anaemia at different treatment times in patients with locally advanced head and neck cancer treated with surgery and postoperative radiotherapy. *Clin Oncol* 2007;**19**:228–33
- Manoochehri Khoshinani H, Afshar S, Najafi R. Hypoxia: a double-edged sword in cancer therapy. *Cancer Invest* 2016;**34**:536–45
- Hughes VS, Wiggins JM, Siemann DW. Tumor oxygenation and cancer therapy-then and now. *Br J Radiol* 2019;**92**:1–12
- Noman MZ, Hasmim M, Messai Y, Terry S, Kieda C, Janji B, Chouaib S. Hypoxia: a key player in antitumor immune response. A review in the theme: cellular responses to hypoxia. *Am J Physiol Cell Physiol* 2015;**309**:C569–79
- Bosco MC, D'Orazi G, Del Bufalo D. Targeting hypoxia in tumor: a new promising therapeutic strategy. *J Exp Clin Cancer Res* 2020;**39**:8–10
- Jahanban-Esfahlan R, de la Guardia M, Ahmadi D, Yousefi B. Modulating tumor hypoxia by nanomedicine for effective cancer therapy. *J Cell Physiol* 2018;**233**:2019–31
- Doguchi H, Saio M, Kuniyoshi S, Matsuzaki A, Yoshimi N. The enhancing effects of hyperbaric oxygen on mouse skin carcinogenesis. *J Toxicol Pathol* 2014;**27**:67–72
- Qian X, Zhang Q, Shao N, Shan Z, Cheang T, Zhang Z, Su Q, Wang S, Lin Y. Respiratory hyperoxia reverses immunosuppression by regulating myeloid-derived suppressor cells and PD-L1 expression in a triple-negative breast cancer mouse model. *Am J Cancer Res* 2019;**9**:529–45
- Yttersian Sletta K, Tveitaras MK, Lu N, Engelsen AST, Reed RK, Garmann-Johnsen A, Stuhr L. Oxygen-dependent regulation of tumor growth and metastasis in human breast cancer xenografts. *PLoS One* 2017;**12**:e0183254
- Lee HY, Kim IK, Lee HI, Lee HY, Kang HS, Yeo CD, Kang HH, Moon HS, Lee SH. Combination of carboplatin and intermittent normobaric hyperoxia synergistically suppresses benzo[a]pyrene-induced lung cancer. *Korean J Intern Med* 2018;**33**:541–51
- Kim SW, Kim IK, Ha JH, Yeo CD, Kang HH, Kim JW, Lee SH. Normobaric hyperoxia inhibits the progression of lung cancer by inducing apoptosis. *Exp Biol Med* 2018;**243**:739–48
- FDA. *Hyperbaric oxygen therapy: don't be misled*, www.fda.gov/consumers/consumer-updates/hyperbaric-oxygen-therapy-dont-be-misled (2013, accessed 10 April 2020)
- Sherman LF, Bacon HE. An evaluation of oxygen therapy as an adjunct in the postoperative management of carcinoma of the rectum and sigmoid colon. *Am J Surg* 1951;**81**:105–10
- Moen I, Tronstad KJ, Kolmannskog O, Salvesen GS, Reed RK, Stuhr LE. Hyperoxia increases the uptake of 5-fluorouracil in mammary tumors independently of changes in interstitial fluid pressure and tumor stroma. *BMC Cancer* 2009;**9**:446–54
- Kawasoe Y, Yokouchi M, Ueno Y, Iwaya H, Yoshida H, Komiya S. Hyperbaric oxygen as a chemotherapy adjuvant in the treatment of osteosarcoma. *Oncol Rep* 2009;**22**:1045–50
- Selvendiran K, Kuppasamy ML, Ahmed S, Bratasz A, Meenakshisundaram G, Rivera BK, Khan M, Kuppasamy P. Oxygenation inhibits ovarian tumor growth by downregulating STAT3 and cyclin-D1 expressions. *Cancer Biol Ther* 2010;**10**:386–90
- Sun S, Lee D, Lee NP, Pu JK, Wong ST, Lui WM, Fung CF, Leung GK. Hyperoxia resensitizes chemoresistant human glioblastoma cells to temozolomide. *J Neurooncol* 2012;**109**:467–75
- Lee D, Sun S, Ho AS, Kiang KM, Zhang XQ, Xu FF, Leung GK. Hyperoxia resensitizes chemoresistant glioblastoma cells to temozolomide through unfolded protein response. *Anticancer Res* 2014;**34**:2957–66
- Hatfield SM, Kjaergaard J, Lukashev D, Schreiber TH, Belikoff B, Abbott R, Sethumadhavan S, Philbrook P, Ko K, Cannici R, Thayer M, Rodig S, Kutok JL, Jackson EK, Karger B, Podack ER, Ohta A, Sitkovsky MV. Immunological mechanisms of the antitumor effects of supplemental oxygenation. *Sci Transl Med* 2015;**7**:1–12
- Benito J, Ramirez MS, Millward NZ, Velez J, Harutyunyan KG, Lu H, Shi YX, Matre P, Jacamo R, Ma H, Konoplev S, McQueen T, Volgin A, Protopopova M, Mu H, Lee J, Bhattacharya PK, Marszalek JR, Davis RE, Bankson JA, Cortes JE, Hart CP, Andreoff M, Konopleva M. Hypoxia-Activated prodrug TH-302 targets hypoxic bone marrow niches in preclinical leukemia models. *Clin Cancer Res* 2016;**22**:1687–98
- Nuongo M, Brigida AL, Mascolo L, Gaudino G. Possible therapeutic effects of ozone mixture on hypoxia in tumor development. *Anticancer Res* 2017;**37**:425–35
- Stepien K, Ostrowski RP, Matyja E. Hyperbaric oxygen as an adjunctive therapy in treatment of malignancies, including brain tumours. *Med Oncol* 2016;**33**:101–10
- Tibbles PM, Edelsberg JS. Hyperbaric-oxygen therapy. *N Engl J Med* 1996;**334**:1642–8
- Weaver LK, Howe S. Normobaric measurement of arterial oxygen tension in subjects exposed to hyperbaric oxygen. *Chest* 1992;**102**:1175–81
- Beppu T, Kamada K, Yoshida Y, Arai H, Ogasawara K, Ogawa A. Change of oxygen pressure in glioblastoma tissue under various conditions. *J Neurooncol* 2002;**58**:47–52
- Heyboer M 3rd, Sharma D, Santiago W, McCulloch N. Hyperbaric oxygen therapy: side effects defined and quantified. *Adv Wound Care* 2017;**6**:210–24
- Raa A, Stansberg C, Steen VM, Bjerkvig R, Reed RK, Stuhr LE. Hyperoxia retards growth and induces apoptosis and loss of glands and blood vessels in DMBA-induced rat mammary tumors. *BMC Cancer* 2007;**7**:23–32
- Braks JA, Spiegelberg L, Koljenovic S, Ridwan Y, Keereweer S, Kanaar R, Wolvius EB, Essers J. Optical imaging of tumor response to hyperbaric oxygen treatment and irradiation in an orthotopic mouse model of head and neck squamous cell carcinoma. *Mol Imaging Biol* 2015;**17**:633–42
- Rockswold SB, Rockswold GL, Zaun DA, Zhang X, Cerra CE, Bergman TA, Liu J. A prospective, randomized clinical trial to compare the effect of hyperbaric to normobaric hyperoxia on cerebral metabolism, intracranial pressure, and oxygen toxicity in severe traumatic brain injury. *J Neurosurg* 2010;**112**:1080–94

42. Ivanov KP, Sokolova IB, Vovenko EP. Oxygen transport in the rat brain cortex at normobaric hyperoxia. *Eur J Appl Physiol Occup Physiol* 1999;**80**:582-7
43. Braun RD, Lanzen JL, Snyder SA, Dewhirst MW. Comparison of tumor and normal tissue oxygen tension measurements using OxyLite or microelectrodes in rodents. *Am J Physiol Heart Circ Physiol* 2001;**280**:H2533-44
44. Stuhr LE, Raa A, Oyan AM, Kalland KH, Sakariassen PO, Petersen K, Bjerkvig R, Reed RK. Hyperoxia Retards growth and induces apoptosis, changes in vascular density and gene expression in transplanted gliomas in nude rats. *J Neurooncol* 2007;**85**:191-202
45. Le QT, Courter D. Clinical biomarkers for hypoxia targeting. *Cancer Metastasis Rev* 2008;**27**:351-62
46. Masoud GN, Li W. HIF-1 α pathway: role, regulation and intervention for cancer therapy. *Acta Pharm Sin B* 2015;**5**:378-89
47. Semenza GL, Wang GL. A nuclear factor induced by hypoxia via de novo protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. *Mol Cell Biol* 1992;**12**:5447-54
48. Lopez-Barneo J, Simon MC. Cellular adaptation to oxygen deficiency beyond the Nobel award. *Nat Commun* 2020;**11**:607-9
49. Majmundar AJ, Wong WJ, Simon MC. Hypoxia-inducible factors and the response to hypoxic stress. *Mol Cell* 2010;**40**:294-309
50. Dengler VL, Galbraith M, Espinosa JM. Transcriptional regulation by hypoxia inducible factors. *Crit Rev Biochem Mol Biol* 2014;**49**:1-15
51. Koyasu S, Kobayashi M, Goto Y, Hiraoka M, Harada H. Regulatory mechanisms of hypoxia-inducible factor 1 activity: two decades of knowledge. *Cancer Sci* 2018;**109**:560-71
52. Harada H. Hypoxia-inducible factor 1-mediated characteristic features of cancer cells for tumor radioresistance. *J Radiat Res* 2016;**57**:i99-i105
53. Xia Y, Jiang L, Zhong T. The role of HIF-1 α in chemo-/radioresistant tumors. *Oncotargets Ther* 2018;**11**:3003-11
54. Sethumadhavan S, Silva M, Philbrook P, Nguyen T, Hatfield SM, Ohta A, Sitkovsky MV. Hypoxia and hypoxia-inducible factor (HIF) down-regulate antigen-presenting MHC class I molecules limiting tumor cell recognition by T cells. *PLoS One* 2017;**12**:e0187314
55. Lv Y, Zhao S, Han J, Zheng L, Yang Z, Zhao L. Hypoxia-inducible factor-1 α induces multidrug resistance protein in colon cancer. *Oncotargets Ther* 2015;**8**:1941-8
56. Hussein AA, Forouzanfar T, Bloemena E, de Visscher J, Brakenhoff RH, Leemans CR, Helder MN. A review of the most promising biomarkers for early diagnosis and prognosis prediction of tongue squamous cell carcinoma. *Br J Cancer* 2018;**119**:724-36
57. Fallah J, Rini BI. HIF inhibitors: status of current clinical development. *Curr Oncol Rep* 2019;**21**:6-15
58. Hatfield SM, Kjaergaard J, Lukashev D, Belikoff B, Schreiber TH, Sethumadhavan S, Abbott R, Philbrook P, Thayer M, Shujia D, Rodig S, Kutok JL, Ren J, Ohta A, Podack ER, Karger B, Jackson EK, Sitkovsky M. Systemic oxygenation weakens the hypoxia and hypoxia inducible factor 1 α -dependent and extracellular adenosine-mediated tumor protection. *J Mol Med* 2014;**92**:1283-92
59. Wang Y, Yin K, Tian J, Xia X, Ma J, Tang X, Xu H, Wang S. Granulocytic Myeloid-Derived suppressor cells promote the stemness of colorectal cancer cells through exosomal S100A9. *Adv Sci* 2019;**6**:1901278
60. Das U. A radical approach to cancer. *Med Sci Monit* 2002;**8**:Ra79-92
61. Allen RG, Balin AK. Effects of oxygen on the antioxidant responses of normal and transformed cells. *Exp Cell Res* 2003;**289**:307-16
62. Folkman J. Seminars in medicine of the Beth Israel hospital, boston. Clinical applications of research on angiogenesis. *N Engl J Med* 1995;**333**:1757-63
63. Chen S, Sang N. Hypoxia-inducible factor-1: a critical player in the survival strategy of stressed cells. *J Cell Biochem* 2016;**117**:267-78
64. Wang GL, Jiang BH, Rue EA, Semenza GL. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O₂ tension. *Proc Natl Acad Sci USA* 1995;**92**:5510-4
65. Gatenby RA, Gillies RJ. Why do cancers have high aerobic glycolysis? *Nat Rev Cancer* 2004;**4**:891-9
66. Mahon PC, Hirota K, Semenza GL. FIH-1: a novel protein that interacts with HIF-1 α and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev* 2001;**15**:2675-86
67. Terraneo L, Virgili E, Caretti A, Bianciardi P, Samaja M. In vivo hyperoxia induces hypoxia-inducible factor-1 α overexpression in LNCaP tumors without affecting the tumor growth rate. *Int J Biochem Cell Biol* 2014;**51**:65-74
68. Rocco M, D'Itri L, D, Bels D, Corazza F, Balestra C. The "normobaric oxygen paradox": a new tool for the anesthetist? *Minerva Anestesiol* 2014;**80**:366-72
69. Chowdhury AK, Watkins T, Parinandi NL, Saatian B, Kleinberg ME, Usatyuk PV, Natarajan V. Src-mediated tyrosine phosphorylation of p47phox in hyperoxia-induced activation of NADPH oxidase and generation of reactive oxygen species in lung endothelial cells. *J Biol Chem* 2005;**280**:20700-11
70. Kim JY, Lee JY. Targeting tumor adaption to chronic hypoxia: implications for drug resistance, and how it can be overcome. *Int J Mol Sci* 2017;**18**:1-13
71. Clark JM, Lambertsen CJ. Pulmonary oxygen toxicity: a review. *Pharmacol Rev* 1971;**23**:37-133
72. Kallet RH, Matthay MA. Hyperoxic acute lung injury. *Respir Care* 2013;**58**:123-41
73. Clark JM, Lambertsen CJ, Gelfand R, Flores ND, Pisarello JB, Rossman MD, Elias JA. Effects of prolonged oxygen exposure at 1.5, 2.0, or 2.5 ATA on pulmonary function in men (predictive studies V). *J Appl Physiol* 1999;**86**:243-59
74. Ciarlone GE, Hinojo CM, Stavitzski NM, Dean JB. CNS function and dysfunction during exposure to hyperbaric oxygen in operational and clinical settings. *Redox Biol* 2019;**27**:101159
75. McMonnies CW. Hyperbaric oxygen therapy and the possibility of ocular complications or contraindications. *Clin Exp Optom* 2015;**98**:122-5
76. He CH, Waxman AB, Lee CG, Link H, Rabach ME, Ma B, Chen Q, Zhu Z, Zhong M, Nakayama K, Nakayama KI, Homer R, Elias JA. Bcl-2-related protein A1 is an endogenous and cytokine-stimulated mediator of cytoprotection in hyperoxic acute lung injury. *J Clin Invest* 2005;**115**:1039-48
77. Zhou Q, Huang G, Yu X, Xu W. A novel approach to estimate ROS origination by hyperbaric oxygen exposure, targeted probes and specific inhibitors. *Cell Physiol Biochem* 2018;**47**:1800-8
78. Körpınar Ş, Uzun H. The effects of hyperbaric oxygen at different pressures on oxidative stress and antioxidant status in rats. *Medicina* 2019;**55**:205-12
79. Sanders LH, Timothy Greenamyre J. Oxidative damage to macromolecules in human parkinson disease and the rotenone model. *Free Radic Biol Med* 2013;**62**:111-20
80. Stadtman ER, Levine RL. Protein oxidation. *Ann N Y Acad Sci* 2000;**899**:191-208
81. Fruhwirth GO, Hermetter A. Mediation of apoptosis by oxidized phospholipids. *Subcell Biochem* 2008;**49**:351-67
82. Auten RL, Whorton MH, Nicholas Mason S. Blocking neutrophil influx reduces DNA damage in hyperoxia-exposed newborn rat lung. *Am J Respir Cell Mol Biol* 2002;**26**:391-7
83. Barazzone C, White CW. Mechanisms of cell injury and death in hyperoxia: role of cytokines and bcl-2 family proteins. *Am J Respir Cell Mol Biol* 2000;**22**:517-9
84. Panayiotidis MI, Rancourt RC, Allen CB, Riddle SR, Schneider BK, Ahmad S, White CW. Hyperoxia-induced DNA damage causes decreased DNA methylation in human lung epithelial-like A549 cells. *Antioxid Redox Signal* 2004;**6**:129-36
85. Gore A, Muralidhar M, Espey MG, Degenhardt K, Mantell LL. Hyperoxia sensing: from molecular mechanisms to significance in disease. *J Immunotoxicol* 2010;**7**:239-54
86. Budinger GR, Tso M, McClintock DS, Dean DA, Sznajder JI, Chandel NS. Hyperoxia-induced apoptosis does not require mitochondrial reactive oxygen species and is regulated by bcl-2 proteins. *J Biol Chem* 2002;**277**:15654-60

87. Sue RD, Belperio JA, Burdick MD, Murray LA, Xue YY, Dy MC, Kwon JJ, Keane MP, Strieter RM. CXCR2 is critical to hyperoxia-induced lung injury. *J Immunol* 2004;**172**:3860–8
88. Murray LA, Knight DA, McAlonan L, Argentieri R, Joshi A, Shaheen F, Cunningham M, Alexopolou L, Flavell RA, Sarisky RT, Hogaboam CM. Deleterious role of TLR3 during hyperoxia-induced acute lung injury. *Am J Respir Crit Care Med* 2008;**178**:1227–37
89. Plafki C, Peters P, Almeling M, Welslau W, Busch R. Complications and side effects of hyperbaric oxygen therapy. *Aviat Space Environ Med* 2000;**71**:119–24
90. Al-Waili NS, Butler GJ, Beale J, Abdullah MS, Finkelstein M, Mellow M, Rivera R, Petrillo R, Carrey Z, Lee B, Allen M. Influences of hyperbaric oxygen on blood pressure, heart rate and blood glucose levels in patients with diabetes mellitus and hypertension. *Arch Med Res* 2006;**37**:991–7
91. Adamson IY, Young L, Orr FW. Tumor metastasis after hyperoxic injury and repair of the pulmonary endothelium. *Lab Invest* 1987;**57**:71–7
92. Ma Z, Moruzzi N, Catrina SB, Grill V, Bjorklund A. Hyperoxia inhibits glucose-induced insulin secretion and mitochondrial metabolism in rat pancreatic islets. *Biochem Biophys Res Commun* 2014;**443**:223–8
93. Karakashev SV, Reginato MJ. Progress toward overcoming hypoxia-induced resistance to solid tumor therapy. *Cancer Manag Res* 2015;**7**:253–64
94. Erler JT, Cawthorne CJ, Williams KJ, Koritzinsky M, Wouters BG, Wilson C, Miller C, Demonacos C, Stratford IJ, Dive C. Hypoxia-mediated down-regulation of bid and bax in tumors occurs via hypoxia-inducible factor 1-dependent and -independent mechanisms and contributes to drug resistance. *Mol Cell Biol* 2004;**24**:2875–89
95. Ding Z, Yang L, Xie X, Xie F, Pan F, Li J, He J, Liang H. Expression and significance of hypoxia-inducible factor-1 alpha and MDR1/P-glycoprotein in human Colon carcinoma tissue and cells. *J Cancer Res Clin Oncol* 2010;**136**:1697–707
96. Suzuki Y, Tanaka K, Negishi D, Shimizu M, Yoshida Y, Hashimoto T, Yamazaki H. Pharmacokinetic investigation of increased efficacy against malignant gliomas of carboplatin combined with hyperbaric oxygenation. *Neurol Med Chir* 2009;**49**:193–7; discussion 7
97. Stuhr LE, Iversen VV, Straume O, Maehle BO, Reed RK. Hyperbaric oxygen alone or combined with 5-FU attenuates growth of DMBA-induced rat mammary tumors. *Cancer Lett* 2004;**210**:35–40
98. Kindwall EP. *Contraindications and side effects to hyperbaric oxygen treatment*. 2nd ed. Flagstaff, AZ: Best Publishing Company, 2002, pp. 83–99
99. Upton PG, Yamaguchi KT, Myers S, Kidwell TP, Anderson RJ. Effects of antioxidants and hyperbaric oxygen in ameliorating experimental doxorubicin skin toxicity in the rat. *Cancer Treat Rep* 1986;**70**:503–7
100. Goldiner PL, Carlon GC, Cvitkovic E, Schweizer O, Howland WS. Factors influencing postoperative morbidity and mortality in patients treated with bleomycin. *Br Med J* 1978;**1**:1664–7
101. Forman HJ, York JL, Fisher AB. Mechanism for the potentiation of oxygen toxicity by disulfiram. *J Pharmacol Exp Ther* 1980;**212**:452–5
102. Karagoz B, Suleymanoglu S, Uzun G, Bilgi O, Aydinoz S, Haholu A, Turken O, Onem Y, Kandemir EG. Hyperbaric oxygen therapy does not potentiate doxorubicin-induced cardiotoxicity in rats. *Basic Clin Pharmacol Toxicol* 2008;**102**:287–92
103. Torp KD, Carraway MS, Ott MC, Stolp BW, Moon RE, Piantadosi CA, Freiberger JJ. Safe administration of hyperbaric oxygen after bleomycin: a case series of 15 patients. *Undersea Hyperb Med* 2012;**39**:873–9
104. Sia J, Szymd R, Hau E, Gee HE. Molecular mechanisms of Radiation-Induced cancer cell death: a primer. *Front Cell Dev Biol* 2020;**8**:41–8
105. Ward JF. DNA damage produced by ionizing radiation in mammalian cells: identities, mechanisms of formation, and reparability. *Prog Nucleic Acid Res Mol Biol* 1988;**35**:95–125
106. Ruan K, Song G, Ouyang G. Role of hypoxia in the hallmarks of human cancer. *J Cell Biochem* 2009;**107**:1053–62
107. Hsia TC, Yang JS, Chen GW, Chiu TH, Lu HF, Yang MD, Yu FS, Liu KC, Lai KC, Lin CC, Chung JG. The roles of endoplasmic reticulum stress and Ca²⁺ on rhEIN-induced apoptosis in A-549 human lung cancer cells. *Anticancer Res* 2009;**29**:309–18
108. Gray LH, Conger AD, Ebert M, Hornsey S, Scott OC. The concentration of oxygen dissolved in tissues at the time of irradiation as a factor in radiotherapy. *Br J Radiol* 1953;**26**:638–48
109. Watson ER, Halnan KE, Dische S, Saunders MI, Cade IS, McEwen JB, Wiernik G, Perrins DJ, Sutherland I. Hyperbaric oxygen and radiotherapy: a medical research council trial in carcinoma of the cervix. *Br J Radiol* 1978;**51**:879–87
110. Bennett MH, Feldmeier J, Smee R, Milross C. Hyperbaric oxygenation for tumour sensitisation to radiotherapy. *Cochrane Database Syst Rev* 2018;**4**:1–115
111. Chen JR, Xu HZ, Ding JB, Qin ZY. Radiotherapy after hyperbaric oxygenation in malignant gliomas. *Curr Med Res Opin* 2015;**31**:1977–84
112. Rojas A, Carl U, Reghebi K. Effect of normobaric oxygen on tumor radiosensitivity: fractionated studies. *Int J Radiat Oncol Biol Phys* 1990;**18**:547–53
113. Overgaard J. Hypoxic radiosensitization: adored and ignored. *J Clin Oncol* 2007;**25**:4066–74
114. Ohta A. Oxygen-dependent regulation of immune checkpoint mechanisms. *Int Immunol* 2018;**30**:335–43
115. Lequeux A, Noman MZ, Xiao M, Sauvage D, Van Moer K, Viry E, Bocci I, Hasmim M, Bosseler M, Berchem G, Janji B. Impact of hypoxic tumor microenvironment and tumor cell plasticity on the expression of immune checkpoints. *Cancer Lett* 2019;**458**:13–20
116. Multhoff G, Vaupel P. Hypoxia compromises anti-Cancer immune responses. *Adv Exp Med Biol* 2020;**1232**:131–43
117. Synnestvedt K, Furuta GT, Comerford KM, Louis N, Karhausen J, Eltzschig HK, Hansen KR, Thompson LF, Colgan SP. Ecto-5'-nucleotidase (CD73) regulation by hypoxia-inducible factor-1 mediates permeability changes in intestinal epithelia. *J Clin Invest* 2002;**110**:993–1002
118. Eltzschig HK, Thompson LF, Karhausen J, Cotta RJ, Ibla JC, Robson SC, Colgan SP. Endogenous adenosine produced during hypoxia attenuates neutrophil accumulation: coordination by extracellular nucleotide metabolism. *Blood* 2004;**104**:3986–92
119. Hasmim M, Noman MZ, Messai Y, Bordereaux D, Gros G, Baud V, Chouaib S. Cutting edge: hypoxia-induced nanog favors the intratumoral infiltration of regulatory T cells and macrophages via direct regulation of TGF-beta1. *J Immunol* 2013;**191**:5802–6
120. Kunzli BM, Bernlochner MI, Rath S, Kaser S, Csizmadia E, Enyoji K, Cowan P, d'Apice A, Dwyer K, Rosenberg R, Perren A, Friess H, Maurer CA, Robson SC. Impact of CD39 and purinergic signalling on the growth and metastasis of colorectal cancer. *Purinergic Signal* 2011;**7**:231–41
121. Stagg J, Divisekera U, Duret H, Sparwasser T, Teng MW, Darcy PK, Smyth MJ. CD73-deficient mice have increased antitumor immunity and are resistant to experimental metastasis. *Cancer Res* 2011;**71**:2892–900
122. Noman MZ, Desantis G, Janji B, Hasmim M, Karray S, Dessen P, Bronte V, Chouaib S. PD-L1 is a novel direct target of HIF-1alpha, and its blockade under hypoxia enhanced MDSC-mediated T cell activation. *J Exp Med* 2014;**211**:781–90
123. Allard B, Pommey S, Smyth MJ, Stagg J. Targeting CD73 enhances the antitumor activity of anti-PD-1 and anti-CTLA-4 mAbs. *Clin Cancer Res* 2013;**19**:5626–35
124. Ohta A, Kini R, Ohta A, Subramanian M, Madasu M, Sitkovsky M. The development and immunosuppressive functions of CD4(+) CD25(+) FoxP3(+) regulatory T cells are under influence of the adenosine-A2A adenosine receptor pathway. *Front Immunol* 2012;**3**:190–201
125. Corzo CA, Condamine T, Lu L, Cotter MJ, Youn JI, Cheng P, Cho HI, Celis E, Quiceno DG, Padhya T, McCaffrey TV, McCaffrey JC, Gabrilovich DI. HIF-1alpha regulates function and differentiation of myeloid-derived suppressor cells in the tumor microenvironment. *J Exp Med* 2010;**207**:2439–53
126. Ryzhov S, Novitskiy SV, Goldstein AE, Biktasova A, Blackburn MR, Biaggioni I, Dikov MM, Feoktistov I. Adenosinergic regulation of the expansion and immunosuppressive activity of CD11b+Gr1+ cells. *J Immunol* 2011;**187**:6120–9
127. Chiu DK, Tse AP, Xu IM, Di Cui J, Lai RK, Li LL, Koh HY, Tsang FH, Wei LL, Wong CM, Ng IO, Wong CC. Hypoxia inducible factor HIF-1

- promotes myeloid-derived suppressor cells accumulation through ENTPD2/CD39L1 in hepatocellular carcinoma. *Nat Commun* 2017;**8**:517–28
128. Sanina N, Chopenko N, Mazeika A, Davydova L, Leonova G, Stenkova A, Uversky VN, Kostetsky E. Immunogenicity and protective activity of a chimeric protein based on the domain III of the Tick-Borne encephalitis virus E protein and the OmpF porin of *Yersinia pseudotuberculosis* incorporated into the TI-complex. *Int J Mol Sci* 2018;**19**:2988–3001
129. Baran N, Konopleva M. Molecular pathways: hypoxia-activated prodrugs in cancer therapy. *Clin Cancer Res* 2017;**23**:2382–90
130. AiErken N, Shi HJ, Zhou Y, Shao N, Zhang J, Shi Y, Yuan ZY, Lin Y. High PD-L1 expression is closely associated with tumor-infiltrating lymphocytes and leads to good clinical outcomes in chinese triple negative breast cancer patients. *Int J Biol Sci* 2017;**13**:1172–9